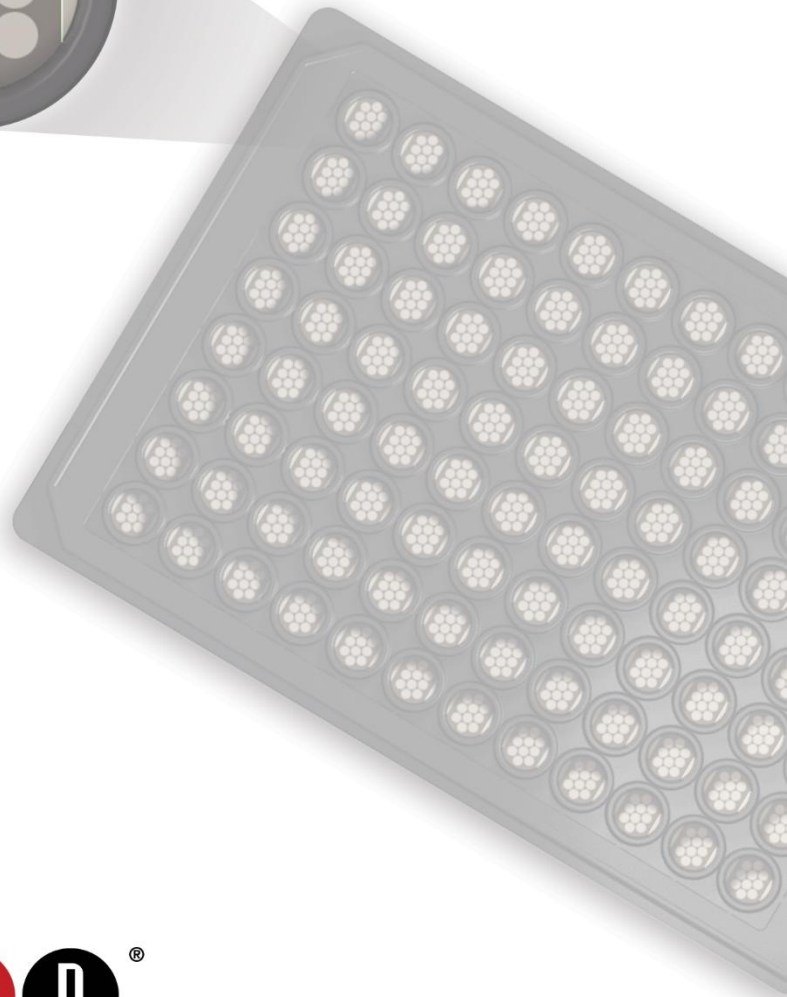


MSD[®] MULTI-SPOT Assay System

Respiratory Panels 5 & 6 Kits

V-PLEX[®]



V-PLEX[®] Respiratory Panels 5 & 6

V-PLEX Respiratory Panels 5 and 6 are multiplex serology assays used to detect IgG antibodies against viral pathogens that cause common upper and lower respiratory infections. These viral pathogens include enterovirus, influenza virus, metapneumovirus, parainfluenza virus, rhinovirus, respiratory syncytial virus, and SARS-CoV-2.

This package insert must be read in its entirety before using this product.

**FOR RESEARCH USE ONLY.
NOT FOR USE IN DIAGNOSTIC PROCEDURES.**

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Introduction

MESO SCALE DISCOVERY® V-PLEX Respiratory Kits measure the amount of anti-IgG antibodies to respiratory pathogens such as enterovirus EV-A71, enterovirus EV-D68, metapneumovirus, respiratory syncytial virus, influenza A (A/Darwin/2021 H3, and A/Wisconsin/2019 H1), influenza B (B/Austria/2021), parainfluenza (serotypes 1, 2, 3, and 4), rhinovirus C, and SARS CoV-2 (BA-5).

Principle of the Assay

The V-PLEX Respiratory Serology Kits quantitatively measure antibodies to antigens from common respiratory viral pathogens (Tables 1 and 2). Plates are provided with antigens on spots in the wells of a 96-well plate (Figure 1). Antibodies in the sample bind to the antigens on the spots and IgG anti-human antibodies conjugated with MSD SULFO-TAG™ label are used for detection. The plate is read on an MSD instrument, which measures the light emitted from the MSD SULFO-TAG.

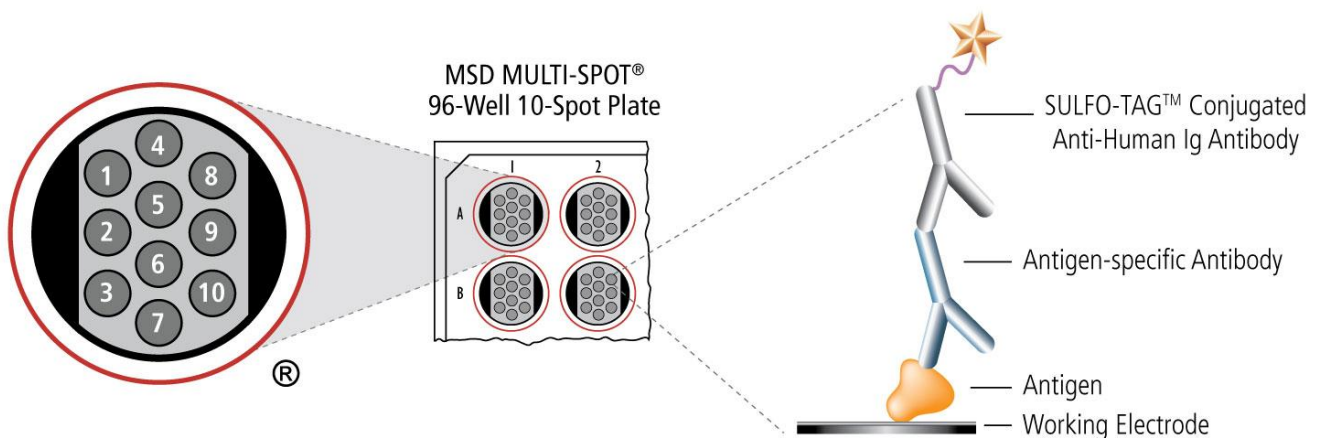


Figure 1. Schematic for V-PLEX Respiratory Serology Kits.

Kit Components

V-PLEX Respiratory Serology Kits are available as panels defined by a set of antigens coated on a 10-spot MULTI-SPOT® 96-well plate. A kit includes a reference standard for quantitation, controls, plate(s), anti-human IgG detection antibody, and all other reagents necessary to conduct the assay.

Tables 1 and 2 describes the available plates and the location of antigens on each plate. Table 3 shows the relationship between the V-PLEX Respiratory Panel 5 and 6 Kits and the plates included in those kits. Table 3 and 4 provide a list of components included in each kit.

Table 1. List of antigens and their spot assignments on the V-PLEX Respiratory Panel 5 Panel

Plate Description	Antigen	Antigen Description
Spot 1	EV-A71 VLP	Enterovirus A71 virus-like particle
Spot 2	—	NA
Spot 3	hMPV F	Metapneumovirus F protein
Spot 4	EV-D68 VLP	Enterovirus D68 virus-like particle
Spot 5	hRV-C VP0	Rhinovirus C VP0
Spot 6	hPIV(1-4) F	Mix of Parainfluenza virus type 1, 2, 3, and 4 F protein
Spot 7	RSV Pre-Fusion F	Respiratory syncytial virus F protein
Spot 8	SARS-CoV-2 Spike (BA.5)	SARS-CoV-2 Omicron BA.5 sublineage spike protein
Spot 9	—	NA
Spot 10	Flu A/B (mix) HA	Mix of Flu A/Wisconsin/2019 H1, Flu A/Darwin/2021 H3, and Flu B/Austria/2021 HA proteins

Table 2. List of antigens and their spot assignments on the V-PLEX Respiratory Panel 6 Panel

Plate Description	Antigen	Antigen Description
Spot 1	Flu A/Wisconsin/2019 H1	Influenza A Hemagglutinin Protein from A/Wisconsin/588/2019 (H1N1)
Spot 2	Flu A/Darwin/2021 H3	Influenza A Hemagglutinin Protein from A/Darwin/6/2021 (H3N2)
Spot 3	Flu B/Austria/2021 HA	Influenza B Hemagglutinin Protein from B/Austria/1359417/2021 (B Victoria Lineage)
Spot 4	—	NA
Spot 5	—	NA
Spot 6	hPIV1 F	Parainfluenza virus type 1 F protein
Spot 7	hPIV2 F	Parainfluenza virus type 2 F protein
Spot 8	hPIV3 F	Parainfluenza virus type 3 F protein
Spot 9	hPIV4 F	Parainfluenza virus type 4 F protein
Spot 10	—	NA

Table 3. Antigen plates included in V-PLEX COVID-19 Serology Kits

Kit	Plate Included
V-PLEX Respiratory Panel 5 Kit	Respiratory Plate 5
V-PLEX Respiratory Panel 6 Kit	Respiratory Plate 6

Table 4. Reagents and Components

Reagent	Storage	Catalog Number	Size	Quantity Supplied	
				5-Plate Kit	25-Plate Kit
MULTI-SPOT 96-Well, 10-Spot plate	2–8 °C	—	10-Spot	5 plates	25 plates
MSD GOLD™ SULFO-TAG Anti-Human IgG Antibody (200X)	2–8 °C	D21APR-3	200 µL	1 vial	5 vials
Diluent 100	2–8 °C	R50AA-2	200 mL	1 bottle	5 bottles
MSD Wash Buffer (20X)	RT	R61AA-1	100 mL	1 bottle	5 bottles
Blocker A	RT	R93BA-2	250 mL	1 bottle	5 bottles
MSD Phosphate Buffer (5X)	RT	R93SA-2	50 mL	1 bottle	5 bottles
MSD GOLD Read Buffer B	RT	R60AM-2	90 mL	1 bottle	5 bottles
Microplate Adhesive Film	RT	—	—	15 sheets	75 sheets
Reference Standard 1	≤-70 °C	C00ADK-2	1 mL	1 vial (IgG assay)	5 vials (IgG assay)
Serology Control Pack 1		C4381-1			
Serology Control 1.1	≤-70 °C		1 mL	1 vial	5 vials
Serology Control 1.2	≤-70 °C		1 mL	1 vial	5 vials
Serology Control 1.3	≤-70 °C		1 mL	1 vial	5 vials

RT = room temperature

Additional Materials and Equipment

- Appropriately sized tubes for reagent preparation
- Deionized water
- 0.2 µM filter needed for Blocker A preparation
- 96-well plates
- Microtiter plate shaker capable of shaking at ~700 rpm
- Microcentrifuge tubes for making serial dilutions
- Automated plate washer or other efficient multi-channel pipetting equipment for washing 96-well plates
- Appropriate liquid handling equipment for desired throughput capable of accurately dispensing 50 µL and 150 µL into a 96-well microplate
- Vortex mixer

Safety

Reference Standard 1 and Serology Control Pack 1 contain human serum and are biosafety level 2 (BSL-2) products. Donors have been tested and found negative for HBsAg, HIV-1 and HIV-2 antibodies, and HCV. Use safe laboratory practices and wear gloves, safety glasses, and laboratory coats when handling kit components. Handle and dispose of all hazardous samples properly in accordance with local, state, and federal guidelines.

Additional product-specific safety information is available in the applicable safety data sheet(s) (SDS), which can be obtained from MSD Customer Service or at www.mesoscale.com[®].

Best Practices

- Mixing or substituting reagents from different sources or different kit lots is not recommended. Lot information is provided in the lot-specific certificate of analysis (COA).
- Assay incubation steps should be performed at 20-26 °C to maximize consistency in signals between runs.
- Avoid prolonged exposure of the detection reagent (stock or diluted) to light. During the antibody incubation step, plates do not need to be shielded from light except for direct sunlight.
- Avoid bubbles in wells at all pipetting steps as they may lead to variable results. Bubbles introduced when adding read buffer may interfere with signal detection.
- Do not touch the pipette tip on the bottom of the wells when pipetting into the MSD plate.
- Use reverse pipetting when necessary to avoid introduction of bubbles. For empty wells, pipette gently to the bottom corner. Do not touch the pipette tip to the bottom of the wells when pipetting into the MSD plate.
- Plate shaking should be vigorous, with a rotary motion between 500-1,000 rpm. Binding reactions may reach equilibrium sooner if shaken in the middle of this range (~700 rpm) or above.
- When performing manual plate washing using a multi-channel pipette, plates should be washed using at least 300 µL of wash buffer per well. Excess residual volume after washing should be removed by gently tapping the plate on a paper towel.
- Do not allow plates to dry after washing steps. Solutions associated with the next assay step should be added to the plate immediately after washing.
- Make sure that the read buffer is at room temperature when adding to the plate.
- To improve interplate precision, keep time intervals consistent between adding read buffer and reading the plate. Unless otherwise directed, read the plate as soon as possible after adding read buffer.
- Do not shake the plate after adding read buffer.
- Remove the plate seals before reading the plate.
- If the sample results are above the top of the calibration curve, dilute the samples, and repeat the assay.
- We do not recommend using a partial plate when running this panel.

Recommended Protocol

Bring all plates and diluents to room temperature. Thaw samples, reference standard, and controls on ice. Thawed reference standard and controls should be equilibrated to room temperature before loading into the plates.

A sample plate layout is shown in Figure 4 (below).

Prepare Blocker A Solution

Follow the preparation procedure in the product insert provided with the Blocker A Kit to prepare the Blocker A solution. You may store unused Blocker A solution according to the instructions in the Blocker A product insert available at www.mesoscale.com.

Prepare Wash Buffer

MSD provides 100 mL of Wash Buffer as a 20X stock solution. Dilute the stock solution before use. PBS + 0.05% Tween-20 can be used as an alternative to MSD Wash Buffer.

For one plate, combine:

- 15 mL of MSD Wash Buffer (20X)
- 285 mL of deionized water

Assay and Antibody Diluent

Use Diluent 100 as assay and antibody diluent.

STEP 1: Prepare Plate

- Remove the plate from its packaging.
- Add 150 μ L/well of Blocker A solution to the plate.
- Seal the plate with an adhesive plate seal and incubate at room temperature with shaking (~700 rpm) for at least 30 minutes.

During this time, prepare calibrators, controls, and samples.

Calibrator Preparation

The kits include a serum-based reference standard, Reference Standard 1, which is used to establish a calibration curve in the assay. The calibration curve is used for calculating the concentration of human IgG against multiple antigens in the V-PLEX Respiratory Serology Kits.

We recommend a 7-point calibration curve with 4-fold serial dilution steps and a zero calibrator blank. Thaw Reference Standard 1 on ice, equilibrate to room temperature, and then add to Diluent 100 to make the calibrator curve solutions.

Appendix A provides assigned values for calibrators in MSD arbitrary units (AU/mL).

Notes:

- For the IgG assays, Reference Standard 1 requires a 10-fold dilution to create the highest calibrator point (CAL-01).

CAL-01 Preparation: Reference Standard 1, 10-fold dilution:

Prepare the highest calibrator solution (CAL-01) by diluting Reference Standard 1 **10-fold**, as shown below (Figure 2):

- ❑ Add 20 μL of the Reference Standard 1 to 180 μL of Diluent 100. Vortex briefly to mix. Label the vial as CAL-01.

CAL-02 to CAL-08 Preparation:

To prepare 7 calibrator solutions for the IgG assays, plus a zero calibrator for up to 2 replicates, perform the following:

- ❑ Prepare the next calibrator (CAL-02) by adding 50 μL of CAL-01 (IgG) to 150 μL of Diluent 100. Vortex briefly to mix.
- ❑ Repeat 4-fold serial dilutions (50 μL previous calibrator into 150 μL Diluent 100) to generate CAL-03 through CAL-07.
- ❑ Use Diluent 100 as the blank (CAL-08).

Note: Stock calibrator is stable for 10 years from the date of manufacture when stored at ≤ -70 °C. The thawed calibrator is stable through five freeze-thaw cycles. Excess diluted calibrator should be discarded after use.

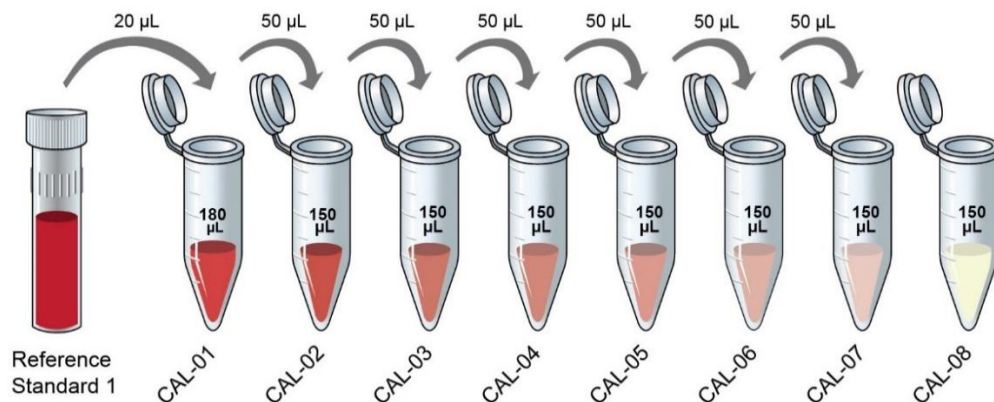


Figure 2. Dilution schema for the preparation of calibrator solutions using a 10-fold dilution of the Reference Standard 1 to generate CAL-01.

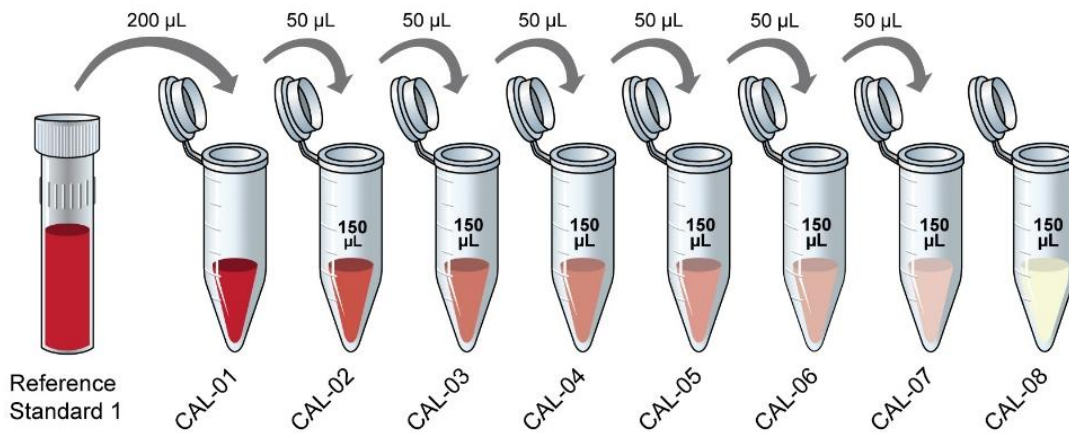


Figure 3. Dilution schema for the preparation of calibrator solutions by using the Reference Standard 1 as neat (undiluted) to generate CAL-01.

Control Preparation

Serology Control Pack 1 consists of three levels of controls, each containing assigned concentrations of human IgG against antigens in the V-PLEX Respiratory Serology Kits. Refer to **Appendix B** for the assigned values in the MSD arbitrary units (AU/mL).

Each control is supplied at the working concentration. Do not dilute prior to use.

Thaw the control on ice and equilibrate to room temperature. Vortex briefly and spin down before loading controls into the plate.

Note: Stock control is stable for 10 years from the date of manufacture when stored at ≤ -70 °C. Thawed control is stable through five freeze-thaw cycles.

Sample Preparation

Prepare the samples by diluting with Diluent 100. The optimal dilution for serum and plasma samples should be determined empirically by the user. Typically, samples are measured at a dilution between 100-fold and 10,000-fold. The lower dilution keeps negative or low samples in the measurable range; higher dilutions prevent saturation of signal with strongly positive samples. This protocol provides guidance for preparing 100-fold and 5,000-fold dilutions, a common choice in vaccine and epidemiological studies.

Note: Saliva and upper respiratory samples are more variable in their composition than serum and plasma samples. Users should run a pilot dilution series to determine the optimal dilution for their saliva and upper respiratory samples.

This protocol provides guidance for preparing both a 100-fold and 5,000-fold diluted sample.

1. To make an intermediate 1:10 dilution in a 2 mL tube, or 96-well plate, combine:
 - 10 µL of sample
 - 90 µL of Diluent 100
2. To make a 1:100 dilution in a 2 mL tube, or 96-well deep well plate, combine:
 - 15 µL of the 1:10 dilution from Step 1.
 - 135 µL of Diluent 100
3. To make a 1:5,000 dilution in a 2 mL tube, or 96-well deep well plate, combine:

10 µL of the 1:100 dilution from Step 2.
490 µL of Diluent 100

STEP 2: Calibrators, Controls, and Sample Addition

After the Blocker A incubation step, wash the plate 3 times with at least 300 µL/well of 1X MSD Wash buffer.

- Add 50 µL/well of diluted samples, calibrators, and controls to the plate.
- Seal the plate with an adhesive plate seal and incubate at room temperature with shaking (~700 rpm) for 2 hours.

During this time, prepare detection antibody solution.

Detection Antibody Solution Preparation

Detection antibody is provided as a 200X stock solution. The working solution is 1X. You will need 6 mL per plate.

To prepare a 1X solution of detection antibody, combine:

- 5,970 µL of Diluent 100
- 30 µL of 200X MSD GOLD SULFO-TAG anti-human IgG Antibody

Note: The MSD GOLD SULFO-TAG Anti-Human IgG Antibody (D21APR-3) is a recombinant monoclonal antibody that binds human immunoglobulin G (IgG), serving as an effective detection reagent in the V-PLEX IgG Serology Assay Kits. Learn more about the MSD GOLD SULFO-TAG Anti-Human IgG Antibody at www.mesoscale.com.

STEP 3: Detection Antibody Addition

After the sample incubation step, wash the plate 3 times with at least 300 µL/well of 1X MSD Wash buffer.

- Add 50 µL/well of 1X detection antibody solution to the plate.
- Seal the plate with an adhesive plate seal and incubate at room temperature with shaking (~700 rpm) for 1 hour.

STEP 4: Read Buffer Addition

After the detection antibody incubation step, wash the plate 3 times with at least 300 µL/well of 1X MSD Wash buffer.

MSD provides MSD GOLD Read Buffer B ready for use. Do not dilute.

- Add 150 µL/well of the MSD GOLD Read Buffer B to the plate.
- Read the plate on the MSD instrument. No incubation in read buffer is required before reading the plate. Read plate immediately after adding read buffer. Do not shake the plate after adding read buffer.

STEP 5: Analysis of Results

Calibration curves used to calculate antibody concentrations are established by fitting the signals from the calibrators to a 4-parameter logistic (or sigmoidal dose-response) model with a $1/Y^2$ weighting. Best quantification of unknown samples is achieved by generating a calibration curve for each plate using a minimum of two replicates at each calibrator level.

Antibody unit concentrations in controls and diluted samples are determined from their ECL signals by backfitting to the calibration curve.

For samples, correcting for dilution provides the final antibody concentrations in undiluted samples (in AU/ml). For example, if 1,000-fold diluted samples are tested, multiply the backfitted concentrations by 1,000.

Controls 1.1, 1.2, and 1.3 are provided pre-diluted for ease of use. Their assigned concentrations reflect the antibody concentrations in the as-provided material. Multiplying the backfitted concentrations of the controls by 5,000 will provide dilution-adjusted concentrations (in AU/mL) that are comparable to concentrations of antibodies in undiluted serum and plasma samples.

Protocol at a Glance

Note: Bring all plates and diluents to room temperature. Thaw samples, reference standard, and controls on ice. Thawed reference standard and controls should be equilibrated to room temperature before loading into the plates.

- Add Blocker A solution; incubate for at least 30 minutes, wash.
- Add samples, calibrators, and controls. Incubate for 2 hours and wash.
- Add Detection Antibody solution. Incubate for 1 hour and wash.
- Add Read Buffer and analyze plate.

Plate Layout

	1	2	3	4	5	6	7	8	9	10	11	12
A	CAL-01		Control 1.1		Sample-06		Sample-14		Sample-22		Sample-30	
B	CAL-02		Control 1.2		Sample-07		Sample-15		Sample-23		Sample-31	
C	CAL-03		Control 1.3		Sample-08		Sample-16		Sample-24		Sample-32	
D	CAL-04		Sample-01		Sample-09		Sample-17		Sample-25		Sample-33	
E	CAL-05		Sample-02		Sample-10		Sample-18		Sample-26		Sample-34	
F	CAL-06		Sample-03		Sample-11		Sample-19		Sample-27		Sample-35	
G	CAL-07		Sample-04		Sample-12		Sample-20		Sample-28		Sample-36	
H	CAL-08		Sample-05		Sample-13		Sample-21		Sample-29		Sample-37	

Figure 4. Sample plate layout that can be used for the assay. Each sample, control, and calibrator is measured in duplicate in side-by-side wells.

Appendix A: Top of the Curve Assignment

The table below shows the assignments for calibrator 1 (CAL-01) for IgG antibodies to antigens in Reference Standard 1. The assignments for Reference Standard 1 (lot number A0080286) differ from previous lots and are presented in a separate table below.

Lot Number A0080286 (Reference Standard 1)

Antigens	Top of the curve assignment for Calibrator 1 (CAL-01)
	IgG MSD AU/mL
SARS-CoV-2 Spike BA.5	11.0
EV-A71 VLP	4.92
EV-D68 VLP	13.8
Flu A/B (mix) HA	87.8
Flu A/Darwin/2021 H3	58.0
Flu A/Wisconsin/2019 H1	95.0
Flu B/Austria/2021 HA	92.0
hMPV F	297
hPIV1 F	43.7
hPIV2 F	45.1
hPIV3 F	128
hPIV4 F	55.7
hRV-C VP0	2.70
hPIV(1-4) F	108
RSV Pre-Fusion F	310

Appendix B: Values for Serology Controls in MSD Arbitrary units (AU/mL)

The tables below show the assigned values for the concentration of IgG antibodies in Serology Control 1.1, Serology Control 1.2, and Serology Control 1.3.

Lot Numbers A00C0825/A00C0826/A00C0827 (Serology Control 1)

Antigens	Concentration of IgG (viald)			Unit of Measure
	Serology Control 1.1	Serology Control 1.2	Serology Control 1.3	
SARS-CoV-2 Spike BA.5	3.15	1.35	0.354	AU/mL
EV-A71 VLP SECTOR	1.40	0.262	NA	AU/mL
EV-A71 VLP QuickPlex Ultra	1.40	0.173	NA	AU/mL
EV-D68 VLP	3.74	1.71	0.156	AU/mL
Flu A/B (mix) HA	25.1	7.17	1.44	AU/mL
Flu A/Darwin/2021 H3	17.4	4.49	0.962	AU/mL
Flu A/Wisconsin/2019 H1	29.3	5.78	2.07	AU/mL
Flu B/Austria/2021 HA	28.6	10.6	1.32	AU/mL
hMPV F	NA	12.7	7.97	AU/mL
hPIV1 F	12.5	2.05	0.821	AU/mL
hPIV2 F	12.9	1.06	0.704	AU/mL
hPIV3 F	34.3	4.88	1.32	AU/mL
hPIV4 F	15.6	4.83	1.32	AU/mL
hRV-C VP0	0.757	0.179	0.0421	AU/mL
hPIV(1-4) F	31.0	4.73	1.44	AU/mL
RSV Pre-Fusion F, SECTOR	91.2	15.7	6.76	AU/mL
RSV Pre-Fusion F, QuickPlex Ultra	NA	15.7	6.76	AU/mL

NA – Not Assigned

Appendix C: Analytical Sensitivity

Limits of quantification (LOQ) were estimated based on the Reference Standard 1 performance over multiple runs. The table below shows the in-well quantitative range for each assay. Multiplying the LLOQ and ULOQ values in the table by the sample dilution factor will provide dilution-adjusted limits of quantification.

Lot Number A0080286 (Reference Standard 1)

Antigens	LLOQ and ULOQ concentrations in MSD arbitrary units (AU/mL)	
	IgG	
	LLOQ	ULOQ
SARS-CoV-2 Spike BA.5	0.0340	11.0
EV-A71 VLP	0.0445	4.92
EV-D68 VLP	0.0560	13.8
Flu A/B (mix) HA	0.0494	87.8
Flu A/Darwin/2021 H3	0.0730	58.0
Flu A/Wisconsin/2019 H1	0.0230	95.0
Flu B/Austria/2021 HA	0.0250	92.0
hMPV F	0.0943	80.0
hPIV1 F	0.0220	43.7
hPIV2 F	0.0628	45.1
hPIV3 F	0.0419	128
hPIV4 F	0.0265	55.7
hRV-C VP0	0.0283	2.70
hPIV(1-4) F	0.0364	108
RSV Pre-Fusion F	0.0600	100

Appendix D: Coated Antigens

Coated Antigens

Antigens	Antigen Description *	Antigen Modifications
SARS-CoV-2 Spike (BA.5)	Severe Acute Respiratory Syndrome Coronavirus 2 Spike Protein Omicron variant BA.5 sublineage	Soluble ectodomain with T4 trimerization domain; C-terminal Strep-Tag and His-Tag
EV-A71 VLP	Enterovirus 71 virus-like particle	N/A
EV-D68 VLP	Enterovirus 68 virus-like particle	N/A
Flu A/B (mix) HA	Mix of Flu A/Wisconsin/2019 H1, Flu A/Darwin/2021 H3, and Flu B/Austria/2021 HA Proteins	Refer to individual antigens
Flu A/Darwin/2021 H3	Influenza A Hemagglutinin Protein from A/Darwin/6/2021 (H3N2)	Soluble ectodomain with T4 trimerization domain; C-terminal His-Tag
Flu A/Wisconsin/2019 H1	Influenza A Hemagglutinin Protein from A/Wisconsin/588/2019 (H1N1)	Soluble ectodomain with T4 trimerization domain; C-terminal His-Tag
Flu B/Austria/2021 HA	Influenza B Hemagglutinin Protein from B/Austria/1359417/2021 (B Victoria Lineage)	Soluble ectodomain with T4 trimerization domain; C-terminal His-Tag
hMPV F	Metapneumovirus F protein	Soluble ectodomain with T4 trimerization domain; C-terminal Strep-Tag and His-Tag
hPIV1 F	Parainfluenza virus type 1 F protein	Soluble ectodomain with GCN4 trimerization domain; C-terminal Strep-Tag and His-Tag
hPIV2 F	Parainfluenza virus type 2 F protein	Soluble ectodomain with GCN4 trimerization domain; C-terminal Strep-Tag and His-Tag
hPIV3 F	Parainfluenza virus type 3 F protein	Soluble ectodomain with GCN4 trimerization domain; C-terminal Strep-Tag and His-Tag
hPIV4 F	Parainfluenza virus type 4 F protein	Soluble ectodomain with GCN4 trimerization domain; C-terminal Strep-Tag and His-Tag
hPIV(1-4) F	Mix of Parainfluenza virus type 1, 2, 3, and 4 F protein	Refer to individual antigens
hRV-C VP0	Rhinovirus C VP0	N-terminal HSA fusion, C-terminal His Tag
RSV Pre-Fusion F	Respiratory Syncytial Virus Pre-Fusion F Protein	Soluble ectodomain with T4 trimerization domain; C-terminal His-Tag and Strep-Tag

*EXPI293 cell line used as an expression system

N/A – Not applicable

Catalog Numbers

Table 5. Catalog Number for V-PLEX Respiratory Serology Kits

Kit Name	IgG	
	5-Plate Kit	25-Plate Kit
Multiplex Kits on the MULTI-SPOT 96-Well, 10-Spot SECTOR plate		
V-PLEX Respiratory Panel 5 Kit	K15754D-2	K15754D-4
V-PLEX Respiratory Panel 6 Kit	K15755D-2	K15755D-4
V-PLEX Respiratory Panel 5 & 6 Kit	K15760D-2	K15760D-4
Multiplex Kits on the 96-Well, 10-Spot plate QuickPlex Ultra Plate		
V-PLEX Respiratory Panel 5 Kit	K15754D-22	K15754D-24
V-PLEX Respiratory Panel 6 Kit	K15755D-22	K15755D-24
V-PLEX Respiratory Panel 5 & 6 Kit	K15760D-22	K15760D-24

Plate Diagram

