U-PLEX® Metabolic Group 1 (mouse)

Singleplex Assays

U-PLEX®

MSD U-PLEX Platform

U-PLEX Metabolic Group 1 (mouse) Singleplex Assays

For use with serum, EDTA plasma, and cell culture supernatants.

This product insert should be read in its entirety before using this product.

FOR RESEARCH USE ONLY.

NOT FOR USE IN DIAGNOSTIC PROCEDURES.

MESO SCALE DISCOVERY®

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Introduction

The U-PLEX Metabolic Group 1 (mouse) contains 58 analytes. Assays for the 12 that are specific to that Group are noted in Table 1. Assays for the other analytes should use the U-PLEX Biomarker Group 1 (mouse) Singleplex Insert. A list of all U-PLEX Assays can be found at *www.mesoscale.com/en/products_and_services/assay_kits/u-plex_gateway*.

There iis a datasheet available for each assay in the U-PLEX portfolio. A representative data set is presented in those datasheets. The datasheets are available at *www.mesoscale.com/support/product_information*.

Table 1. *The 12 assays in U-PLEX Metabolic Group 1 (mouse) that should use this Singleplex Product Insert. Other assays in the Metabolic Group 1 (mouse) should use the Biomarker Group 1 (mouse) Singleplex Insert.*

Assays					
BDNF	FGF-21	Ghrelin (total)	GLP-1 (inactive)	Glucagon	Leptin
C-Peptide	Ghrelin (active)	GLP-1 (active)	GLP-1 (total)	Insulin	PYY (total)

Dash (—) = not applicable

Principle of the Assay

Singleplex assays are supplied on either 96-well or 384-well Streptavidin plates (Figure 1). These plates provide high sensitivity and consistent performance. GOLD-branded plates also deliver excellent inter- and intra-lot uniformity.

Each singleplex assay is supplied with a biotinylated capture antibody that binds to streptavidin on the plate surface. Analytes in the sample bind to the capture antibody. Detection antibodies conjugated with electrochemiluminescent labels (MSD GOLD SULFO TAG™) bind to the analytes to complete the sandwich immunoassay. Once the immunoassay is complete, the plate is loaded into an MSD instrument where a voltage applied to the plate causes the captured labels to emit light. The instrument measures the intensity of emitted light (which is proportional to the amount of analyte present in the sample) and provides a quantitative measure of each analyte in the sample.

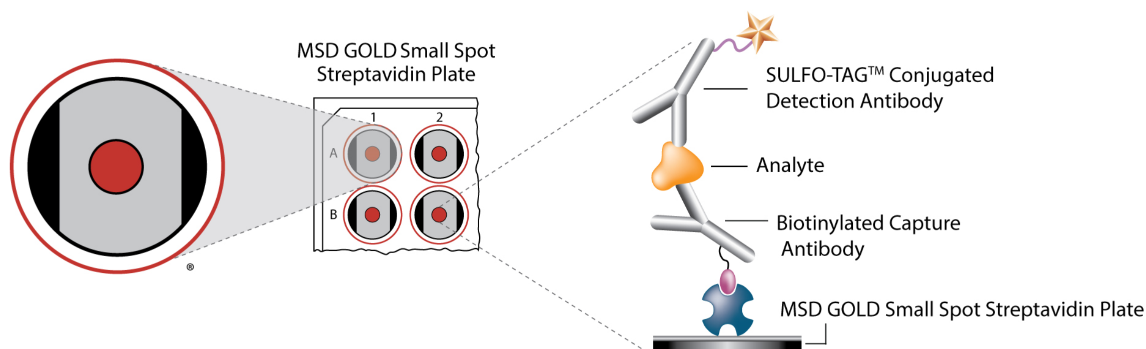


Figure 1. *A U-PLEX singleplex assay on a streptavidin plate.*

Components

Table 2 lists the components provided with U-PLEX Metabolic Group 1 (mouse) Singleplex Assays. U-PLEX singleplex assays are available with either SECTOR™ or QuickPlex 96-well plates.

U-PLEX Singleplex Assays are also available with 384-well SECTOR plates. See Appendix B for details.

Table 2. *Reagents that are supplied with all U-PLEX Metabolic Group 1 (mouse) 96-well Singleplex Assays*

Reagent	Storage	Catalog No.	Size	Quantity Supplied			Description
				1 plate	5 plates	25 plates	
MSD GOLD 96-Well Small Spot Streptavidin SECTOR Plate	2–8 °C	L45SA-1	—	1 plate	5 plates	25 plates	96-well plate, foil sealed, with desiccant.
MSD GOLD 96-Well Small Spot Streptavidin QuickPlex Plate		L4BSA-1					
Diluent 100	2–8 °C	R50AA-4	50 mL	1 bottle	1 bottle	5 bottles	Diluent for capture antibody
Diluent 13	≤–10 °C	R55BB-4	10 mL	1 bottle	—	—	Diluent for samples and calibrator
		R55BB-3	50 mL	—	1 bottle	5 bottles	
Diluent 11	≤–10 °C	R55BA-5	10 mL	1 bottle	—	—	Diluent for detection antibody
		R55BA-3	50 mL	—	1 bottle	5 bottles	
MSD GOLD Read Buffer B	RT	R60AM-1	18 mL	1 bottle	—	—	Buffer to catalyze the electrochemiluminescent reaction
		R60AM-2	90 mL	—	1 bottle	5 bottles	

RT = room temperature
Dash (—) = not applicable

Assay-Specific Reagents

U-PLEX Antibody Set

You will receive a U-PLEX Antibody Set containing a biotinylated capture antibody and a SULFO-TAG conjugated detection antibody (Table 3).

Table 3. *Contents of U-PLEX Antibody Set*

Name	Storage	Size	Quantity Supplied			Description
			1 Plate	5 Plates	25 Plates	
U-PLEX Analyte-Specific Antibody Set	2–8 °C	1-Plate	1	—	—	Set containing biotinylated capture antibody and SULFO-TAG conjugated detection antibody
		5-Plate	—	1	5	

Dash (—) = not applicable

U-PLEX Calibrators

Metabolic Group 1 calibrators (Table 4) are lyophilized multi-analyte blends.

Individual analyte concentrations are provided in the lot-specific certificates of analysis (COA). Assays include one vial of the appropriate Calibrator for each assay plate.

Table 4. Analytes included in the Calibrator blends available for U-PLEX Metabolic Group 1 (mouse)

Name	Storage	Catalog No.	Analytes
Calibrator 18	2–8 °C	C0292-2	BDNF, C-Peptide, FGF-21, Ghrelin, GLP-1 (inactive), GLP-1 (total), PYY (total)
Calibrator 19	2–8 °C	C0293-2	Glucagon, Insulin, Leptin
Ghrelin (active) Calibrator	2–8 °C	C016K-2	Ghrelin (active)
GLP-1 (active) Calibrator	2–8 °C	C016L-2	GLP-1 (active)

Instrument Compatibility

MSD offers U-PLEX assays designed for use on specific instrument platforms (Table 5).

Table 5. Instrument compatibility

Instrument	Assays on 96-well SECTOR plates	Assays on 96-well QuickPlex® plates	Assays on 384-well SECTOR plates
MESO QuickPlex Q 60MM	—	Y	—
MESO® QuickPlex SQ 120	Y	—	—
MESO QuickPlex® SQ 120MM	Y	—	—
MESO SECTOR S 600MM	Y	—	Y
MESO SECTOR® S 600	Y	—	Y

Y = compatible
Dash (—) = not applicable

Additional Materials and Equipment

- ❑ Appropriately sized tubes for reagent preparation
- ❑ Polypropylene tubes for preparing dilutions
- ❑ Liquid-handling equipment suitable for dispensing 10 to 150 µL/well into a 96-well or 384-well microtiter plate
- ❑ Plate-washing equipment: automated plate washer or multichannel pipette
- ❑ Microtiter plate shaker (rotary) capable of shaking at 500–1,000 rpm (1,500 rpm for 384-well plates)
- ❑ MSD Wash Buffer (20X, 100 mL, catalog number R61AA-1) for plate washing. The standard protocol uses a minimum of 200 mL for a 96-well plate and 415 mL for a 384-well plate. Automated plate washers may need overage added to these volumes.
- ❑ DPP-IV Inhibitor. Formulate and store as recommended by the manufacturer
- ❑ MSD Block D-R (catalog number R93BR)
- ❑ Adhesive plate seals
- ❑ Deionized water
- ❑ Vortex mixer
- ❑ MSD Diluent 100 (50 mL, catalog number R50AA-4) may be used to dilute samples that need more than 10-fold dilution.

Safety

Use safe laboratory practices: wear gloves, safety glasses, and lab coats when handling assay components. Handle and dispose of all hazardous samples properly in accordance with local, state, and federal guidelines.

Additional product-specific safety information is available in the applicable safety data sheet (SDS), which can be obtained from MSD Customer Service or at the www.mesoscale.com® website.

Assay Protocol (96-well plates)

Please read the entire detailed Reagent Preparation instructions and the Best Practices (Appendix A) before starting work.

STEP 1: Coat Plates

- ❑ Wash the plate 3 times with at least 150 µL/well of 1X Wash Buffer.
- ❑ Add 200 µL of biotinylated capture antibody to 3.3 mL of Diluent 100. Mix by vortexing.
- ❑ Add 25 µL of the biotinylated antibody solution to each well of the provided 96-well MSD plate. Tap the plate gently on all sides. Seal the plate with an adhesive plate seal and shake for 1 hour at room temperature.
- ❑ Wash the plate 3 times with at least 150 µL/well of 1X MSD Wash Buffer. The plate is now coated and ready for use.

STEP 2: Add Samples and Calibrators

- ❑ Add 50 µL of prepared Calibrator Standard or sample to each well. Seal the plate with an adhesive plate seal. Incubate at room temperature with shaking for 2 hours.

STEP 3: Wash and Add Detection Antibody Solution

- ❑ Wash the plate 3 times with at least 150 µL/well of 1X Wash Buffer.
- ❑ Add 50 µL of detection antibody solution (see Preparation below) to each well. Seal the plate with an adhesive plate seal and incubate at room temperature with shaking for 1 hour.

STEP 4: Wash and Read

- ❑ Wash the plate 3 times with at least 150 µL/well of 1X Wash Buffer.
- ❑ Add 150 µL of MSD GOLD Read Buffer B to each well. Analyze the plate on an MSD instrument. Incubation in Read Buffer is not required before reading the plate.

Important: The BDNF and Insulin assays in particular will experience a time-dependent decrease in signal upon prolonged incubation in Read Buffer. It is recommended that an MSD instrument be prepared to read a plate before adding Read Buffer for these assays.

Reagent Preparation

Important: Upon the first thaw of diluents, aliquot them into suitable volumes before refreezing.

Sample Collection and Handling

Below are general guidelines for sample collection, storage, and handling for metabolic markers. We strongly suggest following these procedures if working with the active forms of protein analytes. If other methods are used, evaluate sample stability under the selected method as needed.

The assay requires 50 µL/well of the sample (25 µL for 384-well plates). Based on the number of replicates desired, prepare an adequate volume of the sample.

Sample Collection (preferred method): Samples should be collected using the BD P800 Collection and Preservation System, which contains a DPP-IV and other protease inhibitor cocktails (Product Number 366420 or 366421). The alternative collection method described below with K_2EDTA tubes can also be used.

Non-P800 collection method: Collect blood in BD Vacutainer K_2EDTA Tubes (Product Number 367841 or 366643). **Immediately** add a DPP-IV inhibitor (0.1 mM final concentration, not provided) and aprotinin (1,000 KIU/mL final concentration) and mix to avoid cleavage/degradation of metabolic peptides.

For BD tubes, process as follows:

- ❑ In a swing-out rotor centrifuge, spin the blood collection tubes as follows;
  - For 2 mL tubes —10 minutes at 1,000 × g (2–8 °C).
  - For 8.5 and 10 mL tubes—20 minutes at 1,300 × g (2–8 °C).
- ❑ Use the plasma immediately or the samples can be stored at 2–8 °C if used within 3 hours. For future use, aliquot the plasma and freeze in suitably sized aliquots at ≤–70 °C.

For samples other than serum and plasma, add a DPP-IV inhibitor (0.1 mM final concentration, not provided) and aprotinin (1,000 KIU/mL final concentration) and use immediately or freeze at ≤–70 °C.

Samples with hemolysis or significant lipemia may hinder accurate measurements.

Repeated freezing and thawing of samples is not recommended. After thawing, centrifuge samples at 2,000 × *g* for 3 minutes to remove particulates before use in the assay. If the samples are clear and no particulates are visible, you may not need to centrifuge. Hold on wet ice or 2–8 °C until processed and used in the assay.

Dilute Samples

Dilute samples two-fold using Metabolic Assay Working Solution. The dilution factor may need to be optimized for the given sample type. Consult MSD technical support if assistance or additional information is required.

Note: For 6CKine/CCL21, BAFF, and NGAL/LCN2, the concentrations in normal serum and EDTA plasma may exceed the standard working range of the assays. Refer to the assay-specific datasheets for additional information.

Prepare Metabolic Assay Working Solution

This solution is used for preparing the calibrator, controls, and diluting the samples.

For one plate, combine the following in a 15 mL tube:

- ❑ 6,965 µL of Assay Diluent
- ❑ 35 µL of aprotinin

Notes:

- Addition of a DPP-IV inhibitor (not provided) to a final concentration of 0.1 mM in the Metabolic Assay Working Solution is strongly recommended. A DPP-IV inhibitor will limit enzymatic action of DPP-IV present in serum/plasma and provides the most accurate measurement of some metabolic analytes.
- **Ghrelin (active) assay only**: Addition of Halt Protease Inhibitor Cocktail, EDTA-Free (Thermo Fisher Scientific, Catalog No. 87785) to a final concentration of 1X in the Metabolic Assay Working Solution is strongly recommended.

Important: Add protease inhibitors immediately before use. Keep the Metabolic Assay Working Solution on ice. Do not freeze the Metabolic Assay Working Solution for later use.

Prepare Detection Antibody Solution

The detection antibody is provided as a 100X stock solution. The working solution is 1X. Prepare the detection antibody solution immediately before use.

For one plate, combine:

- ❑ 60 µL of the supplied 100X detection antibody
- ❑ 5,940 µL of Antibody Diluent

Note, PYY (total) assay only: Addition of MSD Blocker D-R (Catalog No. R93BR, stock concentration 10%) to a final concentration of 0.3% in the Detection Antibody Solution is strongly recommended (180 µL MSD Blocker D-R to a total 6 mL of Detection Antibody Solution). The presence of MSD Blocker D-R does not affect sample quantitation in other assays

Prepare Calibration Standards

Reconstitution

Bring each Calibrator vial to room temperature (see Figure 2; Table 6). Reconstitute lyophilized Calibrators by adding 250 µL of Metabolic Assay Working Solution to the glass vial. This will result in a 10X concentrated stock of the Calibrator. Invert the reconstituted Calibrator at least 3 times. Do not vortex at this point. Let the reconstituted solution equilibrate at room temperature for 15–30 minutes and then vortex briefly. The Calibrator is now ready for use.

Dilutions

The following instructions are for the preparation of 7 Calibrator Standard solutions plus a Zero Calibrator Standard for use in an 8-point standard curve.

Important: Change pipette tips and vortex calibrators after each dilution step. Calibrators are typically run in duplicate. There is sufficient volume of each dilution to run up to six replicates using this process.

- ❑ Prepare Calibrator Standard 1 by adding 25 µL of the reconstituted Calibrator to 225 µL of Metabolic Assay Working Solution. Mix by vortexing.
- ❑ For Calibrator Standard 2, add 75 µL of Calibrator Standard 1 to 225 µL of Metabolic Assay Working Solution.
- ❑ Repeat 4-fold serial dilutions to generate 7 Calibrator Standards. Mix by vortexing between each serial dilution.
- ❑ Use Metabolic Assay Working Solution as Calibrator Standard 8 (zero Calibrator).

Note: For the lot-specific concentration of Calibrators in the blend, refer to the COA supplied with the assay. You can also find a copy of the COA at www.mesoscale.com.

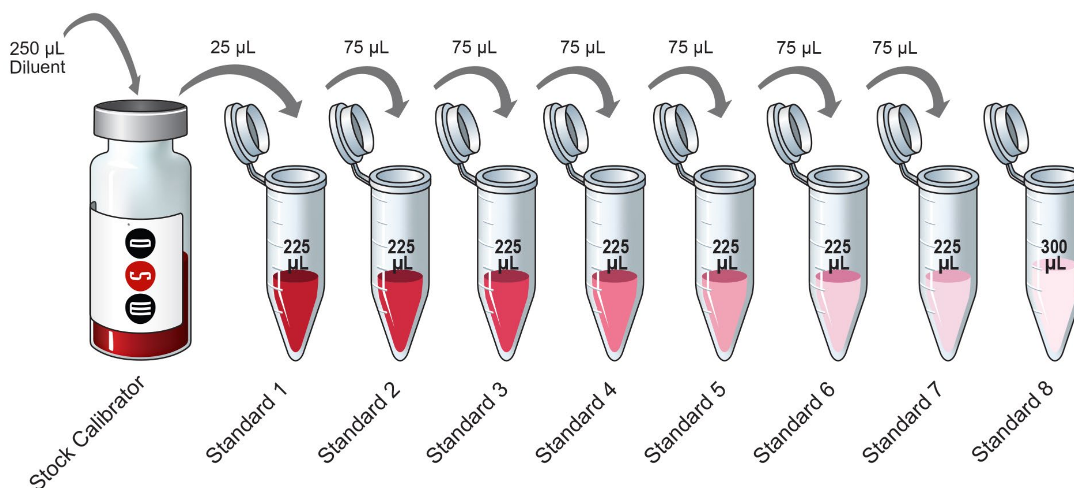


Figure 2. Dilution schema for U-PLEX calibrator standards for singleplex assays.

Table 6. *Serial dilution to generate the standard curve*

Calibrator Standard No.	Tube No.	Source of Calibrator	Volume of Reconstituted Calibrator (µL)	Metabolic Assay Working Solution (µL)	Total Volume (µL)
1	1	Stock Calibrator vial	25	225	250
2	2	Standard 1	75	225	300
3	3	Standard 2	75	225	300
4	4	Standard 3	75	225	300
5	5	Standard 4	75	225	300
6	6	Standard 5	75	225	300
7	7	Standard 6	75	225	300
8 (zero Calibrator)	8	—	0	300	300

Dash (—) = not applicable

Wash Buffer

Prepare a 1X working solution of MSD Wash Buffer (20X, 100 mL, catalog number R61AA-1) by diluting the 20X stock with deionized water. 1X MSD Wash Buffer can be stored at room temperature for up to two weeks. MSD Wash Buffer (20X, 100 mL, catalog number R61AA-1) is ordered separately.

Read Buffer

MSD provides MSD GOLD Read Buffer B ready for use. Do not dilute.

Appendix A

U-PLEX assays are calibrated against a reference Calibrator generated at MSD.

MSD reference Calibrators for the following analytes were evaluated against the NIBSC/WHO International Standards; the ratios of NIBSC/WHO standard relative to MSD Calibrator are shown in the table below. To convert MSD concentrations to biological activity relative to the WHO International Standard, multiply the MSD concentration by the ratio provided (Table 7).

Table 7. Ratios of NIBSC/WHO Standards relative to MSD Calibrators

Analyte	U-PLEX Calibrator	NIBSC/WHO Cat. No.	NIBSC/WHO Unit	MSD Unit	NIBSC/WHO:MSD
BDNF	Calibrator 18	96/534	U/mL	pg/mL	0.0008
Glucagon	Calibrator 19	69/194	IU/mL	pM	3.46×10^{-9}

Calibrator Conversion Factors

For most assays, the Calibrator concentration provided in the COA is in pg/mL. GLP-1 (active, inactive, total) and Glucagon are provided in pM units, and a factor to convert each to pg/mL is provided in Table 8. Insulin is provided in units/mL, and no conversion factor is available.

Table 8. Calibrator concentration conversion factors.

Assay	pM to pg/mL	
GLP-1 (active)	1 pM	3.30 pg/mL
GLP-1 (inactive)	1 pM	3.09 pg/mL
GLP-1 (total)	1 pM	3.09 pg/mL
Glucagon	1 pM	3.48 pg/mL

Notes regarding the following assays

BDNF: The U-PLEX Mouse BDNF Assay has been anchored to NIBSC/WHO standard 96/534. Please see Table 7 for conversion factors.

Insulin, Leptin, C-Peptide, and PYY (total): The U-PLEX Mouse Insulin, Leptin, C-Peptide, and PYY (total) Assays use similar reagents to our previous assays but with different formats and protocols. Each of the U-PLEX Calibrators has been anchored to an internal standard for sustainability. We recommend that users transitioning from previous MSD assays carry out a standard concordance study.

GLP-1 (active) and GLP-1 (total): The U-PLEX GLP-1 (active) and GLP-1 (total) Assays have been developed with better sensitivity and improved specificity compared to our previous GLP-1 assays. The U-PLEX GLP-1 (total) Assay detects both active and inactive fragments of GLP-1 equivalently and enables improved quantitation of total GLP-1 in samples. The Calibrators have been anchored to an internal standard for sustainability. We recommend that users transitioning from previous MSD assays carry out a standard concordance study.

Glucagon: The U-PLEX Glucagon Assay has been anchored to NIBSC/WHO standard 69/194. Please see Table 7 for conversion factors. Compared to previous versions of the assay, the U-PLEX Glucagon Assay has significantly improved sensitivity and dynamic range and enables measurement of native levels of glucagon in 100% of samples collected in P800 tubes. The measured

concentrations of samples may not correlate between the U-PLEX Glucagon Assay and previous MSD glucagon assays. We recommend that users transitioning from previous MSD glucagon assays carry out a standard concordance study.

Alternative Assay Protocols

The suggestions below may be useful for simplifying the protocol.

- ❑ Alternate Protocol 1, Co-incubation: Co-incubating samples and detection antibody solution may improve the sensitivity for some assays. Note that the use of the co-incubation protocol may result in sample concentrations that vary from concentrations obtained with the standard protocol. If this protocol is chosen, we recommend that this protocol be used for the entirety of the research project.
- ❑ Alternate Protocol 2, Shortened Incubation: Some 384-well assays may achieve acceptable performance with shorter incubations. Consider reducing the incubation time of samples in the plate and the detection antibody incubation time.
- ❑ Alternate Protocol 3, Reduced Wash: For cell culture supernatants, you may simplify the protocol by eliminating one of the washes in each step.

Best Practices

- Ensure that all assay components are equilibrated to room temperature before use. Mix well. Bring plates to room temperature before opening the packet.
- Avoid bubbles at each stage of reagent addition because they can lead to variable results. This is very important when adding Read Buffer at the final step prior to reading the plate.
- Plate shaking should be vigorous, with a rotary motion between 500 and 1,000 rpm (1,000 to 1,500 rpm for 384-well plates) depending on the shaker design and orbit. Keep the shaking speed and model the same for long-term studies.
- Tap the plate on a paper towel after washing to ensure the removal of residual fluid.
- Avoid excessive drying of the plate during washing steps, especially if working inside a laminar flow hood or another high-airflow environment. Cover the plate with a new plate seal immediately after washing to protect it from airflow, and add solutions to the plate as soon as possible.
- Use a new adhesive plate seal for all incubation steps. Avoid re-using plate seals.
- Dispense reagents and wash fluids at the side of the well towards the bottom corner.
- Remove the plate seal before reading the plate in the instrument.
- Keep time intervals consistent between the addition of Read Buffer and reading the plate to improve inter-plate precision. Prepare an MSD instrument before adding Read Buffer.
- Do not shake the plate after adding Read Buffer.
- Do not obscure or damage the plate barcode; it is required for the plate reader.
- Only use the reagents provided with this kit.
- Reconstituted or thawed Calibrators should be used immediately. If storage is necessary, divide into suitably sized aliquots, and store immediately at ≤–70 °C.

Appendix B

Components for 384-well plates

Table 7. Reagents that are supplied with all U-PLEX Metabolic Group 1 (mouse) 384-well Singleplex Assays

Reagent	Storage	Catalog No.	Size	Quantity Supplied		Description
				5 Plates	25 Plates	
MSD 384-well Streptavidin SECTOR Plate	2–8 °C	L21SA-1	—	5 plates	25 plates	384-well plate, foil sealed, with desiccant
Diluent 100	2–8 °C	R50AA-4	50 mL	2 bottles	10 bottles	Diluent for biotinylated capture antibody and sample dilution
Diluent 13	≤–10 °C	R55BB-3	50 mL	2 bottles	10 bottles	Diluent for samples and Calibrators
Diluent 11	≤–10 °C	R55BA-3	50 mL	2 bottles	10 bottles	Diluent for detection antibody
MSD GOLD Read Buffer B	RT	R60AM-2	90 mL	1 bottle	5 bottles	Buffer to catalyze the electrochemiluminescent reaction

Dash (—) = not applicable
RT = room temperature

Reagent Preparation for 384-well Plates

Important: Upon the first thaw, aliquot diluents into suitably sized aliquots before refreezing.

Prepare Metabolic Assay Working Solution

This solution is used for preparing the calibrator, controls, and diluting the samples.

For one 384-well plate, combine the following in a 15 mL tube:

- ❑ 13.93 mL of Assay Diluent
- ❑ 70 µL of aprotinin

Notes:

- Addition of a DPPIV inhibitor (not provided) to a final concentration of 0.1 mM in the Metabolic Assay Working Solution is strongly recommended. A DPPIV inhibitor will limit enzymatic action of DPPIV present in serum/plasma and provides the most accurate measurement of some metabolic analytes.
- **Ghrelin (active) assay only**: Addition of Halt Protease Inhibitor Cocktail, EDTA-Free (Thermo Fisher Scientific, Catalog No. 87785) to a final concentration of 1X in the Metabolic Assay Working Solution is strongly recommended.

Important: Protease inhibitors should be added immediately before use. The Metabolic Assay Working Solution should be kept on ice. Do not freeze the Metabolic Assay Working Solution.

Coat 384-well Plate

- ❑ Add 240 µL of biotinylated capture antibody to 11.76 mL of Diluent 100. Mix by vortexing.
- ❑ Add 25 µL of the above solution to each well of the provided plate. Tap the plate gently on all sides. Seal the plate with an adhesive plate seal and incubate with shaking at room temperature for 2 hours.
- ❑ Wash the plate 3 times with 80 µL/well of 1X MSD Wash Buffer. The plate is now coated and ready for use. Plates may be sealed and stored overnight at 4 °C.

Prepare Detection Antibody Solution

The detection antibody is provided as a 100X stock solution. The working solution is 1X. Prepare the detection antibody solution immediately before use.

For one plate, combine:

- ❑ 60 µL of the supplied 100X detection antibody
- ❑ 11.94 mL of Antibody Diluent

Note, PYY (total) assay only: Addition of MSD Blocker D-R (Catalog No. R93BR, stock concentration 10%) to a final concentration of 0.3% in the Detection Antibody Solution is strongly recommended (360 µL MSD Blocker D-R to a total 12 mL of Detection Antibody Solution). The presence of MSD Blocker D-R does not affect sample quantitation in other assays.

Assay Protocol (384-well plates)

Important: Please read the entire detailed Reagent Preparation instructions and the Best Practices (Appendix A) before starting work.

STEP 1: Coat Plates

- ❑ Wash the plate 3 times with at least 80 µL/well of 1X Wash Buffer.
- ❑ Add 240 µL of biotinylated capture antibody to 11.76 mL of Diluent 100. Mix by vortexing.
- ❑ Add 25 µL of the biotinylated antibody solution to each well of the provided 384-well MSD plate. Tap the plate gently on all sides. Seal the plate with an adhesive plate seal and shake for 1 hour at room temperature.
- ❑ Wash the plate 3 times with at least 80 µL/well of 1X MSD Wash Buffer. The plate is now coated and ready for use.

STEP 2: Add Samples and Calibrators

- ❑ Wash the plate 3 times with 80 µL/well of 1X MSD Wash Buffer.
- ❑ Add 25 µL of the prepared Calibrator Standard or sample to each well. Seal the plate with an adhesive plate seal. Incubate at room temperature with shaking for 2 hours.

STEP 3: Wash and Add Detection Antibody Solution

- ❑ Wash the plate 3 times with 80 µL/well of 1X MSD Wash Buffer.
- ❑ Add 25 µL of detection antibody solution to each well. Seal the plate with an adhesive plate seal. Incubate at room temperature with shaking for 2 hours.

STEP 4: Wash and Read

- ❑ Wash the plate 3 times with 80 µL/well of 1X MSD Wash Buffer.
- ❑ Add 40 µL of MSD GOLD Read Buffer B to each well. Analyze the plate on an MSD instrument. Incubation in Read Buffer is not required before reading the plate.

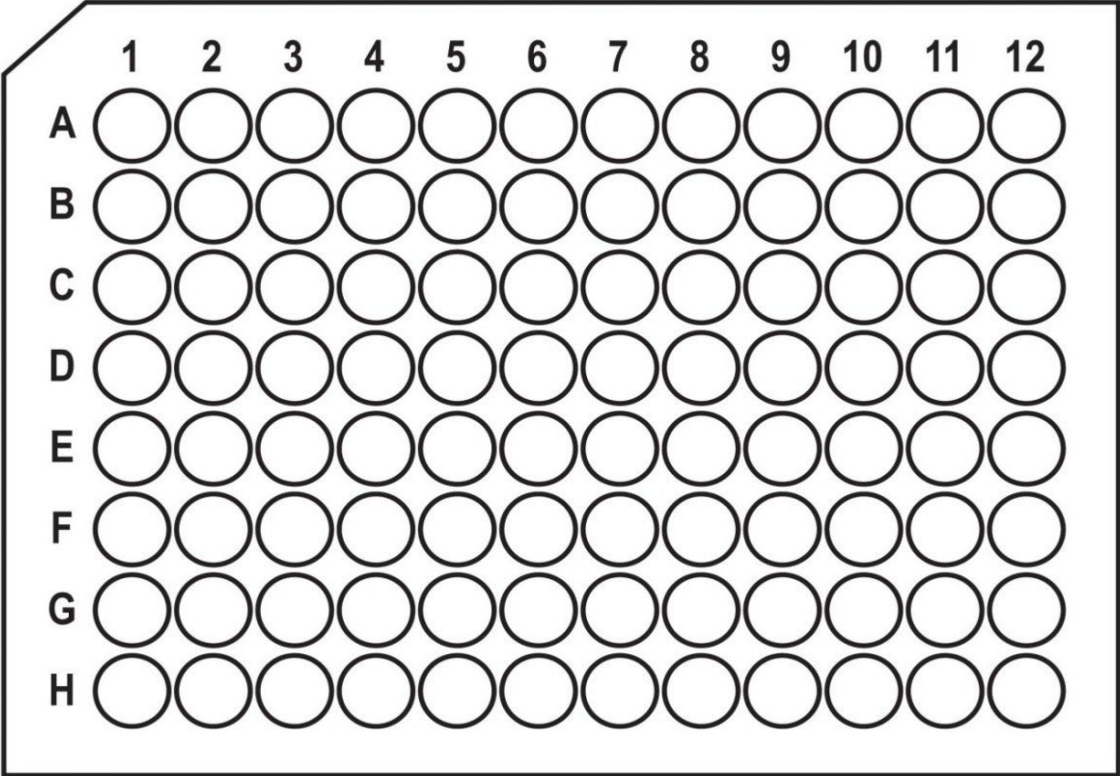
Important: The BDNF and Insulin assays in particular will experience a time-dependent decrease in signal upon prolonged incubation in Read Buffer. It is recommended that an MSD instrument be prepared to read a plate before adding Read Buffer for these assays.

Alternative Assay Protocols

The suggestions below may be useful for simplifying the protocol.

- ❑ **Alternate Protocol, Shortened Incubation:** Some 384-well assays may achieve acceptable performance with shorter incubations. Consider reducing the incubation time of samples in the plate and the detection antibody incubation time.

Plate Diagrams



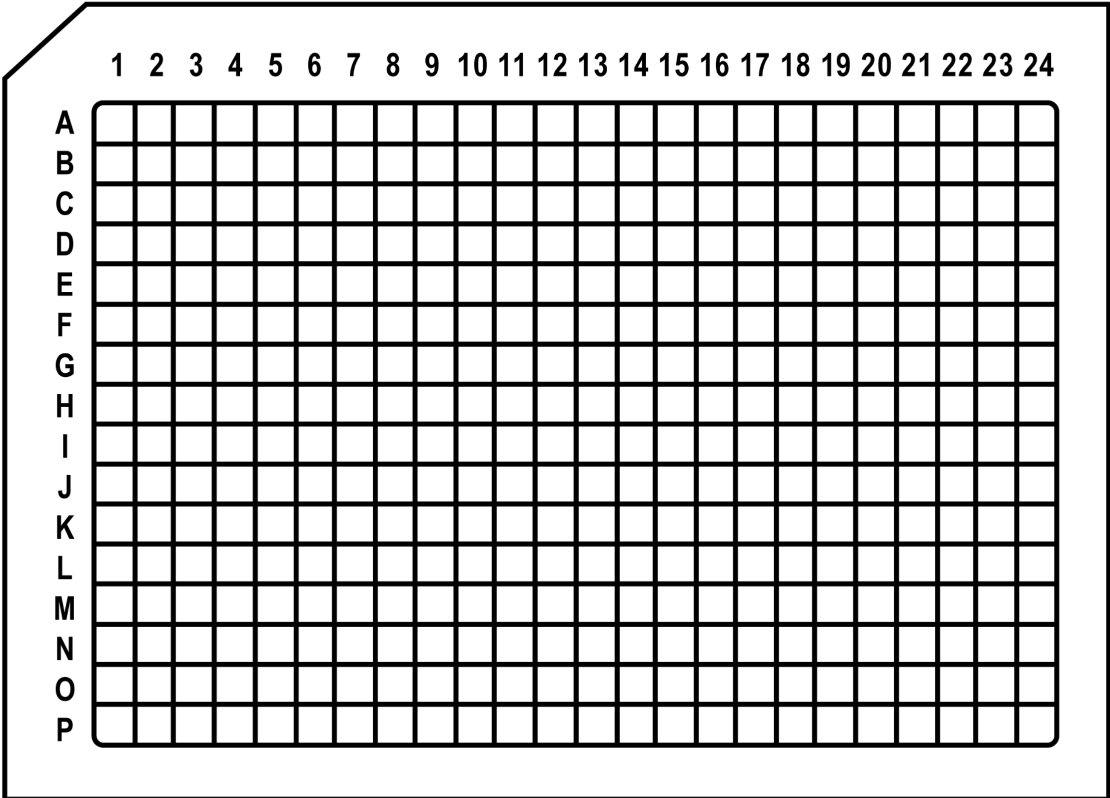


Figure 3. *Plate diagrams. Similar plate layouts can be created in Excel and in the DISCOVERY WORKBENCH® software.*