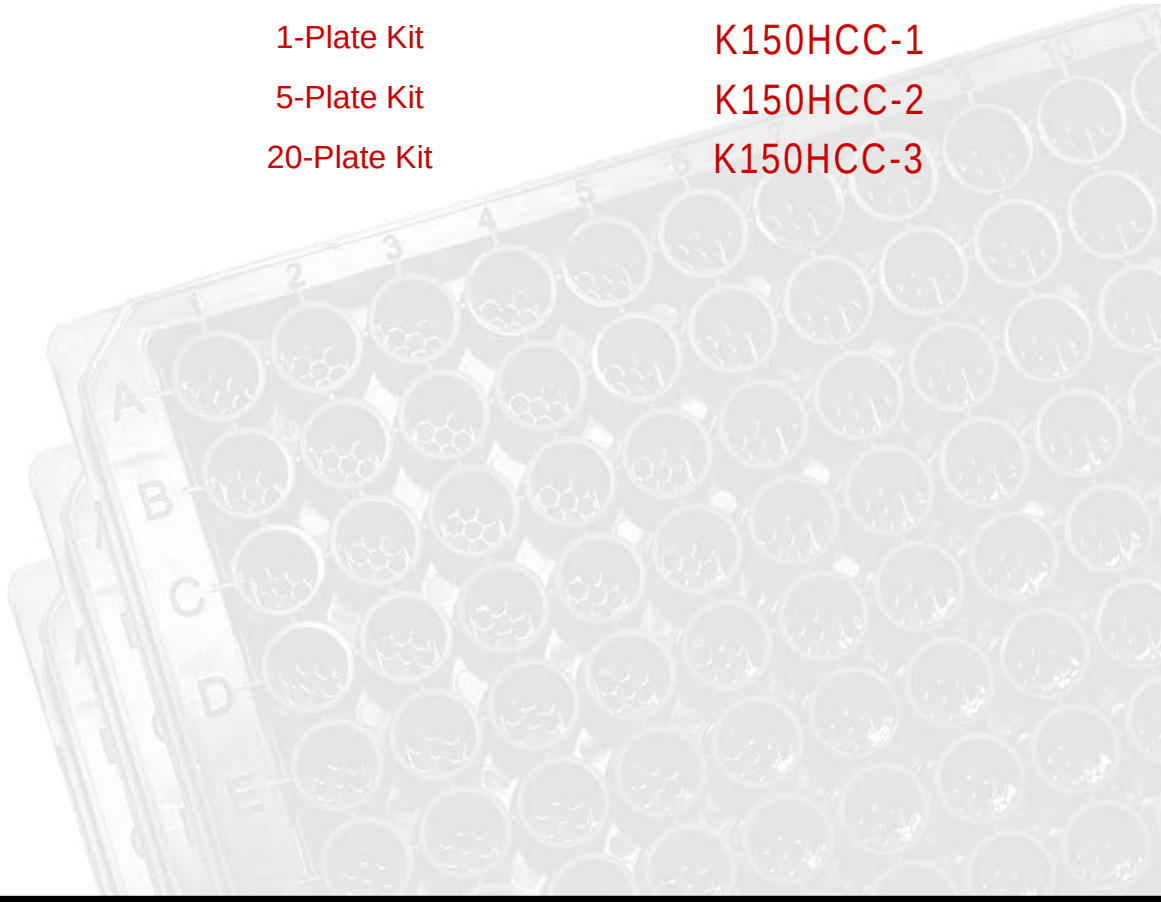
Meso Scale Discovery®

MULTI-ARRAY® Assay System

Mouse/Rat Glucagon Kit

1-Plate Kit	K150HCC-1
5-Plate Kit	K150HCC-2
20-Plate Kit	K150HCC-3



MSD Metabolic Assays

Mouse/Rat Glucagon Kit

This package insert must be read in its entirety before using this product.

FOR RESEARCH USE ONLY.

NOT FOR USE IN DIAGNOSTIC PROCEDURES.

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Ordering Information

ordering information

MSD Customer Service

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Introduction

introduction

Glucagon is a 29-residue polypeptide hormone that is produced in the pancreas by the α-cells of the islets of Langerhans. Glucagon is involved in maintaining normal levels of glucose in the blood by acting on liver glycogen, converting it to glucose. Glucagon is a stimulator of hepatic glycogenolysis, gluconeogenesis, and ketogenesis which are antagonistic effects to those of insulin action, resulting in increased blood glucose levels. Glucagon receptors have been found in liver, kidney, intestinal smooth muscle, brain and adipose tissue.

Principle of the Assay

principle of the assay

MSD® metabolic assays provide rapid and convenient methods for measuring the levels of protein targets within single small-volume samples. The assays are available in both singleplex and multiplex formats. In a singleplex assay, an antibody for a specific protein target is coated on one electrode (or "spot") per well. In a multiplex assay, an array of capture antibodies against different targets is patterned on distinct spots in the same well. Our Mouse/Rat Glucagon Assay detects glucagon in a sandwich immunoassay (Figure 1). MSD provides a plate that has been pre-coated with glucagon antibody. The user adds the sample and a solution containing the labeled detection antibody—anti-glucagon labeled with an electrochemiluminescent compound, MSD SULFO-TAG™ label—over the course of one or more incubation periods. Glucagon in the sample binds to capture antibody immobilized on the working electrode surface; recruitment of the labeled detection antibody by bound analyte completes the sandwich. The user adds an MSD read buffer that provides the appropriate chemical environment for electrochemiluminescence and loads the plate into an MSD SECTOR instrument for analysis. Inside the SECTOR instrument, a voltage applied to the plate electrodes causes the labels bound to the electrode surface to emit light. The instrument measures intensity of emitted light to afford a quantitative measure of glucagon present in the sample.

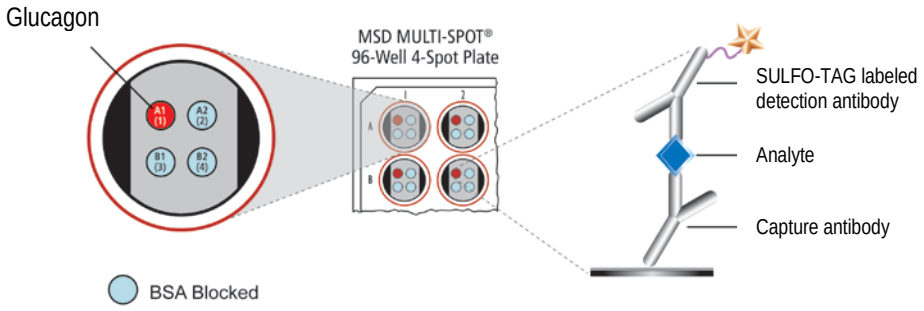


Figure 1. *Sandwich immunoassay on MSD platform. The numbering convention for the different spots is maintained in the software visualization tools, on the plate packaging, and in the data files. Any spot that is not coated with a specific capture antibody is blocked with BSA to reduce non-specific binding to that spot. A unique bar code label on each plate allows complete traceability back to MSD manufacturing records.*

III Reagents Supplied

reagents supplied

Product Description	Storage	Quantity per Kit K150HCC-1	K150HCC-2	K150HCC-3
MULTI-SPOT 96-well Glucagon Plate(s) N450HCA-1	2-8°C	1 plate	5 plates	20 plates
SULFO-TAG Anti-Glucagon Antibody[1] (100X)	2-8°C	1 vial (40 µL)	1 vial (200 µL)	4 vials (200 µL ea)
Glucagon Calibrator 1 µg/mL	≤-70°C	1 vial (15 µL)	5 vials (15 µL ea)	20 vials (15 µL ea)
Blocker A Kit R93AA-2 (250 mL)	RT	1 bottle (250 mL)	1 bottle (250 mL)	4 bottles (250 mL ea)
Aprotinin (200,000 KIU/mL)	2-8°C	1 vial (50 µL)	1 vial (250 µL)	4 vials (250 µL ea)
Blocker D-B (10%)	≤-10°C	1 vial (0.25 mL)	1 vial (1.2 mL)	4 vials (1.2 mL ea)
Diluent 17 R50KA-3 (30 mL)	≤-10°C	1 bottle (30 mL)	2 bottles (30 mL ea)	5 bottles (30 mL ea)
Diluent 100 R50AA-4 (50 mL)	2-8°C	1 bottle (50 mL)	1 bottle (50 mL)	2 bottles (50 mL ea)
Read Buffer T (4X) R92TC-3 (50 mL) R92TC-2 (200 mL)	RT	1 bottle (50 mL)	1 bottle (50 mL)	1 bottle (200 mL)

IV Required Materials and Equipment - not supplied

required materials and equipment — not supplied

- Deionized water for diluting concentrated buffers
- 50 mL tubes for reagent preparation
- 15 mL tubes for reagent preparation
- Polypropylene microcentrifuge tubes for preparing serial dilutions
- Phosphate buffered saline plus 0.05% Tween-20 (PBS-T) for plate washing
- Appropriate liquid handling equipment for desired throughput, capable of dispensing 10 to 150 µL into a 96-well microtiter plate
- Plate washing equipment: automated plate washer or multichannel pipette
- Adhesive plate seals
- Microtiter plate shaker

[1] Some SULFO-TAG labeled detection antibodies may be light-sensitive, so they should be stored in the dark.

V Safety

safety

Safe laboratory practices and personal protective equipment such as gloves, safety glasses, and lab coats should be used at all times during the handling of all kit components. All hazardous samples should be handled and disposed of properly, in accordance with local, state, and federal guidelines.

VI Reagent Preparation

reagent preparation

Bring all reagents to room temperature and thaw the Calibrator stock on ice.
Blocker D-B can tolerate up to 5 freeze-thaw cycles. Alternatively, an aliquot of the blocker can be stored at 2-8°C for up to 1 month.

> **Important:** Upon first thaw, separate Diluent 17 into aliquots appropriate to the size of your assay needs. This diluent can go through up to three freeze-thaw cycles without significantly affecting the performance of the assay.

Prepare Blocker A Solution

Follow instructions included with the Blocker A Kit.

Prepare Metabolic Assay Working Solution

In a 15 mL tube combine (per plate):

- ❑ 40 µL of Aprotinin
- ❑ 7960 µL of Diluent 17

> **Important:** Aprotinin should be added prior to use. The Metabolic Assay Working Solution should be kept on ice. Do not freeze the Metabolic Assay Working Solution for later use.

Prepare Calibrator and Control Solutions

Calibrator for the Mouse/Rat Glucagon Assay is supplied at 1 µg/mL. For the assay, an 8-point standard curve is recommended with 3-fold serial dilution steps and a zero Calibrator. The table below shows the concentrations of the 8-point standard curve:

Standard	Glucagon conc. (pg/mL)	Dilution Factor
Stock Cal. Vial	1000000	
STD-01	10000	100
STD-02	3333	3
STD-03	1111	3
STD-04	370	3
STD-05	123	3
STD-06	41	3
STD-07	14	3
STD-08	0	n/a

To prepare this 8-point standard curve:

1) Prepare the highest Calibrator by transferring 10 µL of the Calibrator stock solution at 1 µg/mL to 990 µL of Metabolic Assay Working Solution.
2) Prepare the next Calibrator by transferring 100 µL of the diluted Calibrator to 200 µL of Metabolic Assay Working Solution. Repeat 3-fold serial dilutions 5 additional times to generate 7 Calibrators.
3) The recommended 8th Standard is Metabolic Assay Working Solution (i.e. zero Calibrator).
4) Diluted Calibrators should be kept on ice prior to addition to the plate.

Note: The standard curve can be modified as necessary to meet specific assay requirements.

Preparation of Serum and Plasma Samples

1) The assay format requires 40 µL of sample per well. An adequate volume of each sample should be prepared depending upon desired number of replicates.
2) There are numerous proteases in serum and plasma that may cause degradation of Glucagon. Blood samples should be drawn into tubes containing 500 KIU Aprotinin per mL of whole blood. Alternately, Aprotinin should be added immediately following blood draw. Invert the blood tube several times to mix the sample.
    a. To obtain serum, tubes containing Aprotinin should be allowed to clot for 30' on a rocker. Spin the tubes for 10 minutes at 1000 x g (4^{o}C) and aliquot serum into separate tubes and store at -80^{o}C until use. Avoid repeated freeze-thaw (> 2) of these aliquots.
    b. Plasma samples should be obtained in vacutainer or syringe containing Na_2EDTA (1.25 mg/mL) and 500 KIU Aprotinin per mL of whole blood. Tubes should be spun for 10 minutes at 1000 x g (4^{o}C) and then plasma immediately aliquotted into separate tubes and stored at -80^{o}C until use. Avoid repeated freeze-thaw (> 2) of these aliquots.
3) Keep isolated or thawed serum/plasma samples on ice or at 4^{o}C prior to subsequent processing or until use in the assay.
4) Samples with hemolysis or significant lipemia may hinder accurate assay measurements.

Prepare Detection Antibody Solution

The Detection Antibody is provided at 100X stock solution. The final concentration of the working Detection Antibody Solution should be at 1X. For each plate used, dilute a 30 µL aliquot of the stock Detection Antibody and 90 µL of 10% Blocker D-B into a final volume of 3 mL of Diluent 100.

Prepare Read Buffer

The Read Buffer should be diluted 4-fold in deionized water to make a final concentration of 1X Read Buffer T. Add 5 mL of 4X Read Buffer T to 15 mL of deionized water for each plate.

Prepare MSD Plate

This plate has been pre-coated with antibody for the analyte shown in Figure 1. The plate can be used as delivered; no additional preparation (e.g., pre-wetting) is required. The plate has also been exposed to a proprietary stabilizing treatment to ensure the integrity and stability of the immobilized antibodies.

VII Assay Protocol

assay protocol

1. **Addition of Blocker A Solution:** Dispense 150 µL of Blocker A Solution into each well. Seal the plate with an adhesive plate seal and incubate for 1 hour with vigorous shaking (300–1000 rpm) at room temperature.
2. **Wash and Addition of Sample or Calibrator:** Wash the plate 3 times with PBS-T. Dispense 20 µL of Metabolic Assay Working Solution into each well of the MSD plate. Immediately add 40 µL of sample or Calibrator into the appropriate wells of the MSD plate. Seal the plate with an adhesive plate seal and incubate for 2 hours with vigorous shaking (300–1000 rpm) at room temperature.
3. **Wash and Addition of the Detection Antibody Solution:** Wash the plate 3 times with PBS-T. Dispense 25 µL of the 1X Detection Antibody Solution into each well of the MSD plate. Seal the plate and incubate for 1 hour with vigorous shaking (300–1000 rpm) at room temperature.
4. **Wash and Read:** Wash the plate 3 times with PBS-T. Add 150 µL of 1X Read Buffer T to each well of the MSD plate. Analyze the plate on the SECTOR Imager. Plates may be read immediately after the addition of Read Buffer.

Notes

Shaking a 96-well MSD MULTI-SPOT plate typically accelerates capture at the working electrode.

Bubbles in the fluid will interfere with reliable reading of MULTI-SPOT plate. Use reverse pipetting techniques to insure bubbles are not created when dispensing the Read Buffer.

VIII Analysis of Results

analysis of results

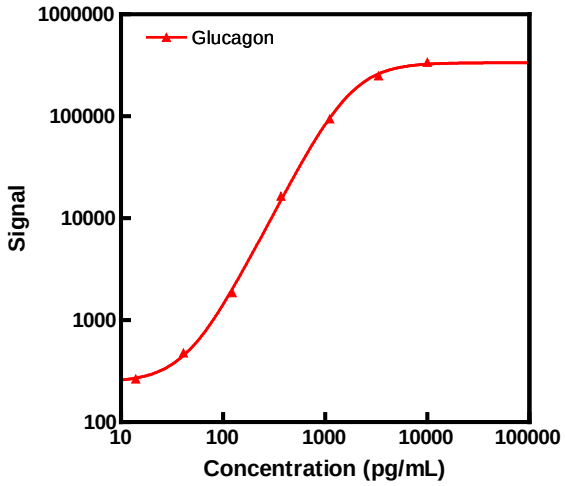
The Calibrators should be run in duplicate to generate a standard curve. The standard curve is modeled using least squares fitting algorithms so that signals from samples with known levels of the analyte of interest can be used to calculate the concentration of analyte in the sample. The assays have a wide dynamic range (3–4 logs) which allows accurate quantification in many samples without the need for dilution. The MSD DISCOVERY WORKBENCH® analysis software utilizes a 4-parameter logistic model (or sigmoidal dose-response) and includes a $1/Y^2$ weighting function. The weighting functionality is important because it provides a better fit of data over a wide dynamic range, particularly at the low end of the standard curve.

IX Typical Standard Curve

typical standard curve

The MSD Mouse/Rat Glucagon Assay is designed for use with mouse and rat serum and plasma samples.

The following standard curve is an example of the dynamic range of the assay. The actual signals may vary. A standard curve should be run for each set of samples and on each plate for the best quantification of unknown samples.



	Glucagon	
Conc. (pg/mL)	**Average Signal**	**%CV**
0	185	13.9
14	265	3.1
41	477	3.3
123	1864	10.8
370	16483	2.7
1111	94445	2.3
3333	249107	8.0
10000	339538	5.3

X Sensitivity

sensitivity

The lower limit of detection (LLOD) is the calculated concentration of the signal that is 2.5 standard deviations over the zero Calibrator. The value below represents the average LLOD over multiple kit lots.

	Glucagon
LLOD (pg/mL)	19

XI Spike Recovery

spike recovery

Serum, EDTA plasma, and heparin plasma were spiked with the Calibrators at multiple values throughout the range of the assay. The Calibrators were spiked into individual mouse samples and therefore spike recovery may depend on specific individual samples. MSD recommends using plasma samples for optimal assay performance.
% Recovery = measured /expected x 100

Sample	Spike Level (pg/mL)	Conc. (pg/mL)	Conc. %CV	% Recovery
Spiked Serum	0	<LLOD	3.1	-
	200	118	2.4	70
	3500	2653	2.5	81
	7000	5264	6.0	87
Spiked EDTA Plasma	0	<LLOD	0.5	-
	200	170	4.8	100
	3500	3088	6.8	94
	7000	6448	3.8	106
Spiked Heparin Plasma	0	<LLOD	4.0	-
	200	140	7.6	82
	3500	2836	6.1	86
	7000	6171	8.0	102

XII Linearity

linearity

Linearity was measured by spiking Calibrator levels in pooled mouse plasma followed by subsequent dilution.
Percent recovery is calculated as the measured concentration divided by the concentration of the previous dilution (expected). % Recovery = measured x dilution factor / expected x 100

Sample	Fold Dilution	Conc. (pg/mL)	Conc. %CV	% Recovery
Serum	1	2195	4.5	100
	2	2652	4.9	121
	4	2685	5.0	101
	8	2379	3.3	89
EDTA Plasma	1	3271	3.2	100
	2	3656	1.9	112
	4	3502	2.2	96
	8	2822	4.6	81
Heparin Plasma	1	2584	4.0	100
	2	3074	3.2	119
	4	2890	2.9	94
	8	2708	5.4	94

XIII Assay Components

assay components

Calibrator	
Analyte	Glucagon
Source	Synthetic human glucagon (amino acids 1-29)

Capture Antibody	
Analyte	Human glucagon
Source	Mouse monoclonal
Isoforms Recognized	n/a
Species cross-reactivity	Human, mouse, rat

Detection Antibody	
Analyte	Human glucagon
Source	Mouse monoclonal, ascites
Isoforms Recognized	Pancreatic glucagon, reacts weakly to gut glucagon
Species cross-reactivity	Human, mouse, rat, sheep, rabbit, pig, canine, pig, guinea pig

XIII References

references

1. Mojsov S, Heinrich G, Wilson IB, Ravazzola M, Orci L, Habener JF. Preproglucagon gene expression in pancreas and intestine diversifies at the level of post-translational processing. J Biol Chem. 1986 Sep 5;261(25):11880-9.

2. Witt S, Dietz H, Ziegler B, Keilacker H, Ziegler M. Production and use of monoclonal glucagon and insulin antibodies--reduction of pancreatic insulin in rats by treatment with complete Freund's adjuvant. Acta Histochem Suppl. 1988;35:217-23

3. Ahrén B. Glucagon secretion in relation to insulin sensitivity in healthy subjects. Diabetologia. 2006 Jan;49(1):117-22

Summary Protocol

MSD 96-well MULTI-ARRAY Mouse/Rat Glucagon Kit

MSD provides this summary protocol for your convenience.
Please read the entire detailed protocol prior to performing the Mouse/Rat Glucagon Assay.

Step 1 : Sample and Reagent Preparation

Bring all reagents to room temperature and thaw the Calibrator stock on ice.

Prepare Blocker A Solution.

Prepare serum or plasma samples.

Prepare Metabolic Assay Working Solution and keep on ice.

Prepare an 8-point standard curve using supplied Calibrator:

- The Calibrator should be diluted in Metabolic Assay Working Solution.
- Dilute the stock Calibrator 1:100 as indicated in Reagent Preparation section, then perform a series of 3-fold dilution steps and a no Calibrator blank.
- Diluted Calibrators should be kept on ice until use.

Note: *The standard curve can be modified as necessary to meet specific assay requirements.*

Prepare Detection Antibody Solution by diluting the 100X Anti-Glucagon Antibody to 1X and the 10% Blocker D-B to 0.3% in 3.0 mL of Diluent 100 per plate.

Prepare 20 mL of 1X Read Buffer T by diluting 4X Read Buffer T with deionized water.

Step 2 : Add Blocker A Solution

Dispense 150 µL/well Blocker A Solution.

Incubate at room temperature with vigorous shaking (300–1000 rpm) for 1 hour.

Step 3 : Wash and Add Sample or Calibrator

Wash plate 3 times with PBS-T.

Dispense 20 µL/well Metabolic Assay Working Solution.

Immediately, dispense 40 µL/well Calibrator or Sample.

Incubate at room temperature with vigorous shaking (300–1000 rpm) for 2 hours.

Step 4 : Wash and Add Detection Antibody Solution

Wash plate 3 times with PBS-T.

Dispense 25 µL/well 1X Detection Antibody Solution.

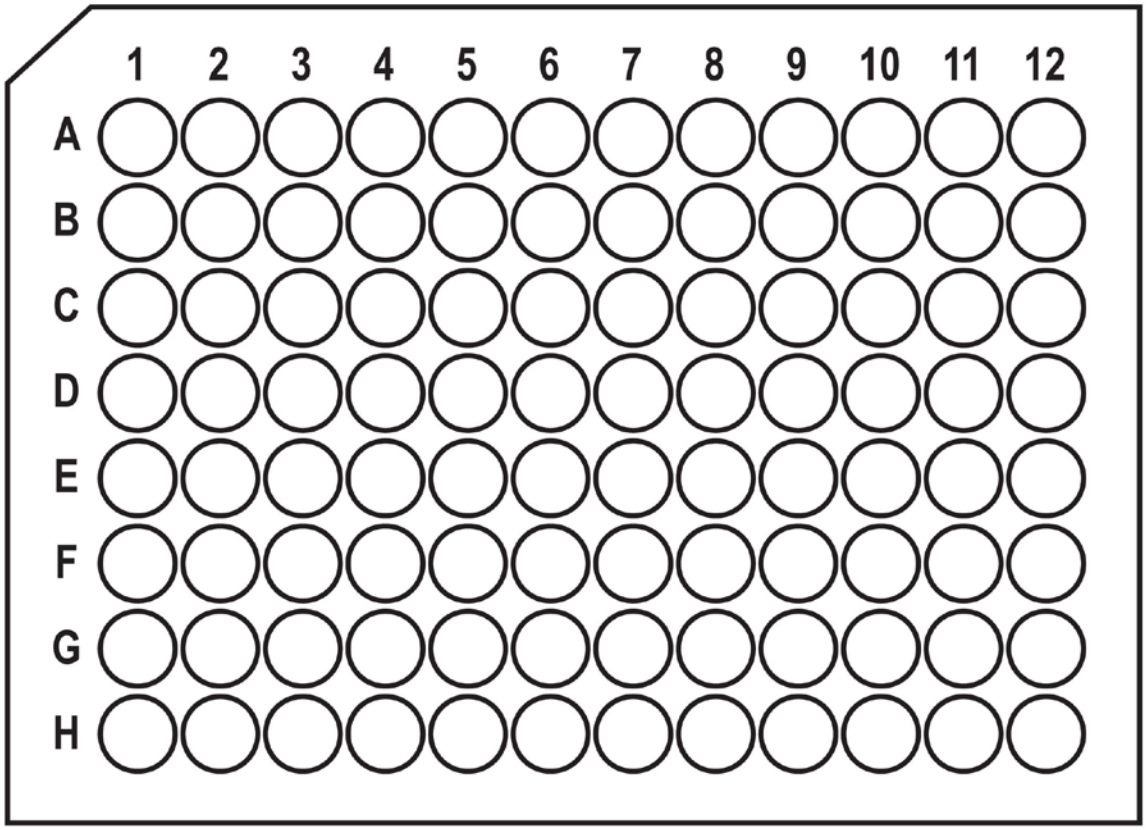
Incubate at room temperature with vigorous shaking (300–1000 rpm) for 1 hour.

Step 5 : Wash and Read Plate

Wash plate 3 times with PBS-T.

Dispense 150 µL/well 1X Read Buffer T.

Analyze plate on SECTOR instrument.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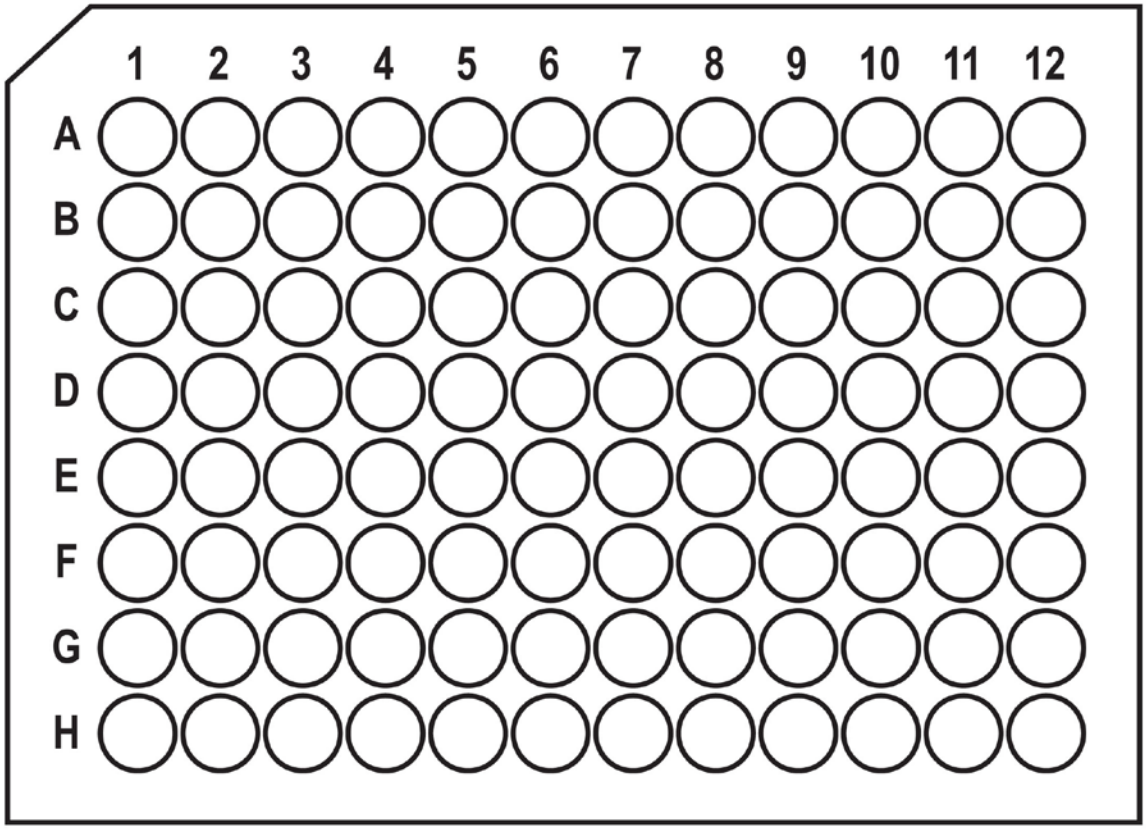
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