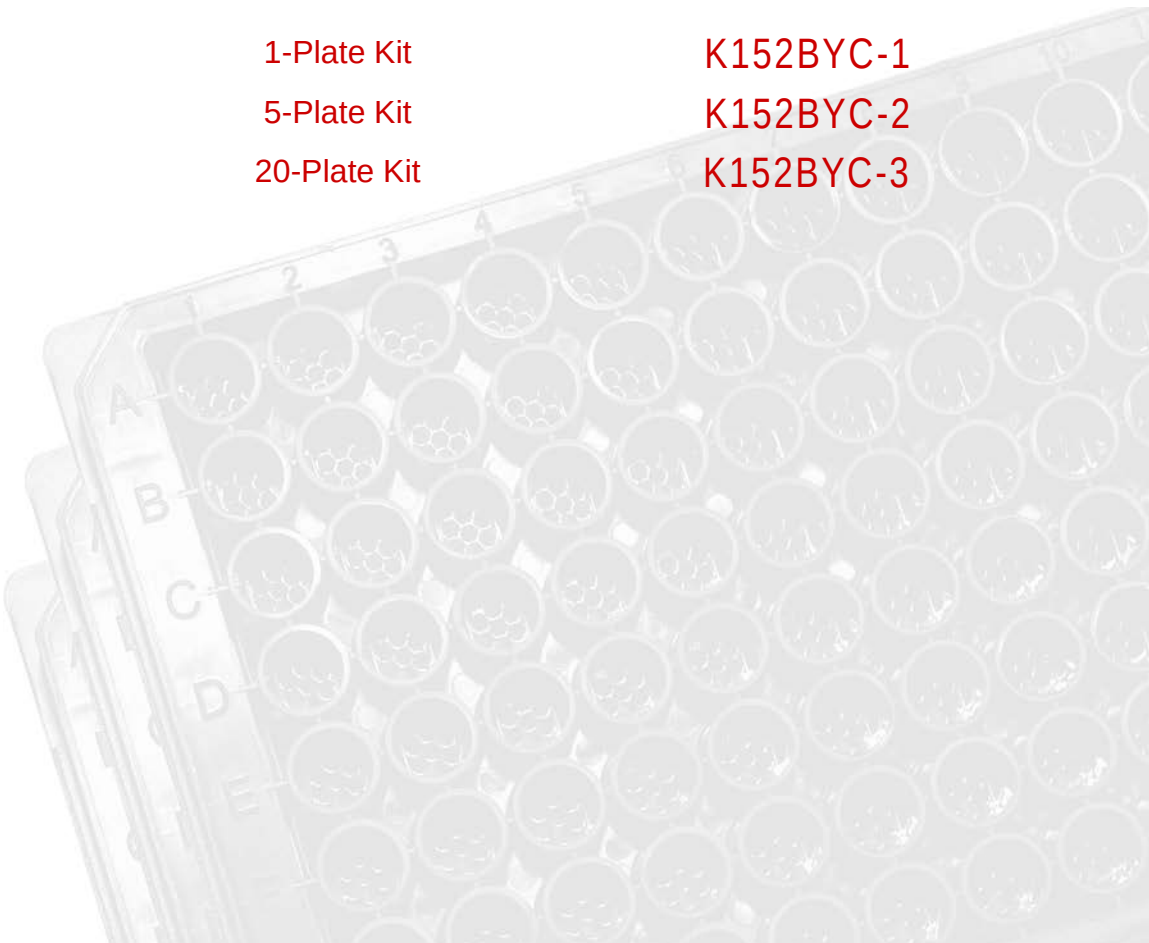
Meso Scale Discovery®

MULTI-ARRAY® Assay System

Mouse Leptin Kit

1-Plate Kit	K152BYC-1
5-Plate Kit	K152BYC-2
20-Plate Kit	K152BYC-3



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MSD Metabolic Assays

Mouse Leptin Kit

This package insert must be read in its entirety before using this product.

FOR RESEARCH USE ONLY.

NOT FOR USE IN DIAGNOSTIC PROCEDURES.

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Table of Contents

table of contents

I. Introduction 4
II. Principle of the Assay 4
III. Reagents Supplied 5
IV. Required Material and Equipment – not supplied 5
V. Safety 5
VI. Reagent Preparation 6
VII. Assay Protocol 7
VIII. Analysis of Results 7
IX. Typical Standard Curve 8
X. Sensitivity 8
XI. Spike Recovery 9
XII. Linearity 9
XIII. Assay Components 10
XIV. References 10
Summary Protocol 12
Plate Diagrams 14

Ordering Information

ordering information

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Introduction

introduction

Leptin is a 16 kD product of the ob gene that is produced and released by adipocytes. Acting via cytokine-like receptors in the CNS, leptin plays a key role in metabolism and regulation of adipose tissue. Leptin is released in amounts mirroring overall body fat stores and acts on neurons and hypothalamic receptors thereby influencing the brain's perception of nutritional energy status and appetite. The absence of functional leptin (or its receptor) leads to uncontrolled food intake and resulting obesity. Fasting reduces circulating insulin and leptin levels in plasma. Leptin may therefore be a critical regulator of obesity often accompanied by insulin resistance and hyperinsulinemia.

Principle of the Assay

principle of the assay

MSD® metabolic assays provide rapid and convenient methods for measuring the levels of protein targets within single small-volume samples. The assays are available in both singleplex and multiplex formats. In a singleplex assay, an antibody for a specific protein target is coated on one electrode (or "spot") per well. In a multiplex assay, an array of capture antibodies against different targets is patterned on distinct spots in the same well. Our Mouse Leptin Assay detects leptin in a sandwich immunoassay (Figure 1). MSD provides a plate that has been pre-coated with leptin antibody. The user adds the sample and a solution containing the labeled detection antibody—anti-leptin labeled with an electrochemiluminescent compound, MSD SULFO-TAG™ label—over the course of one or more incubation periods. Leptin in the sample binds to capture antibody immobilized on the working electrode surface; recruitment of the labeled detection antibody by bound analyte completes the sandwich. The user adds an MSD read buffer that provides the appropriate chemical environment for electrochemiluminescence and loads the plate into an MSD SECTOR instrument for analysis. Inside the SECTOR instrument, a voltage applied to the plate electrodes causes the labels bound to the electrode surface to emit light. The instrument measures intensity of emitted light to afford a quantitative measure of leptin present in the sample.

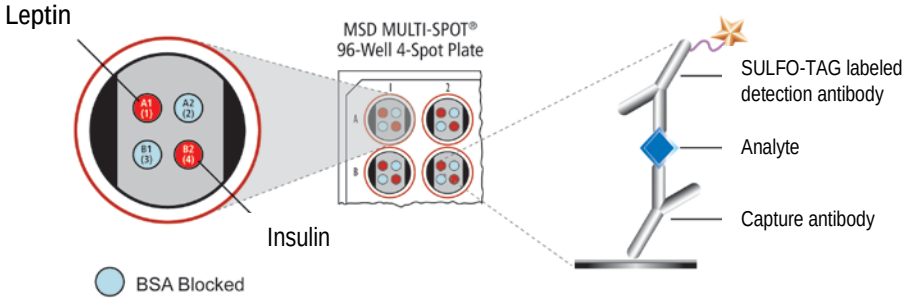


Figure 1. *Sandwich immunoassay on MSD platform. The numbering convention for the different spots is maintained in the software visualization tools, on the plate packaging, and in the data files. Any spot that is not coated with a specific capture antibody is blocked with BSA to reduce non-specific binding to that spot. A unique bar code label on each plate allows complete traceability back to MSD manufacturing records.*

III Reagents Supplied

reagents supplied

Product Description	Storage	Quantity per Kit K152BYC-1	K152BYC-2	K152BYC-3
MULTI-SPOT 96-well Mouse Metabolic Plate(s) N45124A-1	2-8°C	1 plate	5 plates	20 plates
SULFO-TAG Anti-mLeptin Antibody[1] (100X)	2-8°C	1 vial (40 µL)	1 vial (200 µL)	4 vials (200 µL ea)
Mouse Leptin Calibrator 10 µg/mL	≤-70°C	1 vial (15 µL)	5 vials (15 µL ea)	20 vials (15 µL ea)
Blocker A Kit R93AA-2 (250 mL)	RT	1 bottle (250 mL)	1 bottle (250 mL)	4 bottles (250 mL ea)
Diluent 17 R50KA-4 (6 mL) R50KA-3 (30 mL)	≤-10°C	1 bottle (6 mL)	1 bottle (30 mL)	4 bottles (30 mL ea)
Diluent 100 R50AA-4 (50 mL) R50AA-2 (200 mL)	2-8°C	1 bottle (50 mL)	1 bottle (50 mL)	1 bottle (200 mL)
Read Buffer T (4X) R92TC-3 (50 mL) R92TC-2 (200 mL)	RT	1 bottle (50 mL)	1 bottle (50 mL)	1 bottle (200 mL)

IV Required Materials and Equipment - not supplied

required materials and equipment — not supplied

- Deionized water for diluting concentrated buffers
- 50 mL tubes for reagent preparation
- 15 mL tubes for reagent preparation
- Microcentrifuge tubes for preparing serial dilutions
- Phosphate buffered saline plus 0.05% Tween-20 (PBS-T) for plate washing
- Appropriate liquid handling equipment for desired throughput, capable of dispensing 10 to 150 µL into a 96-well microtiter plate
- Plate washing equipment: automated plate washer or multichannel pipette
- Adhesive plate seals
- Microtiter plate shaker

V Safety

safety

Safe laboratory practices and personal protective equipment such as gloves, safety glasses, and lab coats should be used at all times during the handling of all kit components. All hazardous samples should be handled and disposed of properly, in accordance with local, state, and federal guidelines.

[1] Some SULFO-TAG labeled detection antibodies may be light-sensitive, so they should be stored in the dark.

VI Reagent Preparation

reagent preparation

Bring all reagents to room temperature and thaw the Calibrator stock on ice.

> **Important:** Upon first thaw, separate Diluent 17 into aliquots appropriate to the size of your assay needs. This diluent can go through up to three freeze-thaw cycles without significantly affecting the performance of the assay.

Prepare Blocker A Solution

Follow instructions included with the Blocker A Kit.

Prepare Calibrator and Control Solutions

Calibrator for the Mouse Leptin Assay is supplied at 10 µg/mL. For the assay, an 8-point standard curve is recommended with 3-fold serial dilution steps and a zero Calibrator. The table below shows the concentrations of the 8-point standard curve:

Standard	Leptin conc. (pg/mL)	Dilution Factor
Stock Cal. Vial	10000000	
STD-01	100000	100
STD-02	33333	3
STD-03	11111	3
STD-04	3704	3
STD-05	1235	3
STD-06	412	3
STD-07	137	3
STD-08	0	n/a

To prepare this 8-point standard curve:

1) Prepare the highest Calibrator by transferring 10 µL of the Calibrator stock vial to 990 µL of Diluent 100.
2) Prepare the next Calibrator by transferring 100 µL of the diluted Calibrator to 200 µL of Diluent 100. Repeat 3-fold serial dilutions 5 additional times to generate 7 Calibrators.
3) The recommended 8th Standard is Diluent 100 (i.e. zero Calibrator).
4) Diluted Calibrators should be kept on ice prior to addition to the plate.

Note: The standard curve can be modified as necessary to meet specific assay requirements.

Preparation of Serum and Plasma Samples

The assay format requires 10 µL of sample per well. An adequate volume of each sample should be prepared depending upon desired number of replicates.

Prepare Detection Antibody Solution

The Detection Antibody is provided at 100X stock solution. The final concentration of the working Detection Antibody Solution should be at 1X. For each plate used, dilute a 50 µL aliquot of the stock Detection Antibody into 4.95 mL of Diluent 17.

Prepare Read Buffer

The Read Buffer should be diluted 4-fold in deionized water to make a final concentration of 1X Read Buffer T. Add 5 mL of 4X Read Buffer T to 15 mL of deionized water for each plate.

Prepare MSD Plate

This plate has been pre-coated with antibodies for the analytes shown in Figure 1. The plate can be used as delivered; no additional preparation (e.g., pre-wetting) is required. The plate has also been exposed to a proprietary stabilizing treatment to ensure the integrity and stability of the immobilized antibodies.

VII Assay Protocol

assay protocol

1. **Addition of Blocker A Solution:** Dispense 150 µL of Blocker A Solution into each well. Seal the plate with an adhesive plate seal and incubate for 1 hour with vigorous shaking (300–1000 rpm) at room temperature.
2. **Wash and Addition of Sample or Calibrator:** Wash the plate 3 times with PBS-T. Dispense 40 µL of 1X Detection Antibody Solution into each well of the MSD plate. Immediately add 10 µL of sample or Calibrator into the appropriate wells of the MSD plate. Seal the plate with an adhesive plate seal and incubate for 2 hours with vigorous shaking (300–1000 rpm) at room temperature.
3. **Wash and Read:** Wash the plate 3 times with PBS-T. Add 150 µL of 1X Read Buffer T to each well of the MSD plate. Analyze the plate on the SECTOR Imager. Plates may be read immediately after the addition of Read Buffer.

Notes

Shaking a 96-well MSD MULTI-SPOT plate typically accelerates capture at the working electrode.

Bubbles in the fluid will interfere with reliable reading of MULTI-SPOT plate. Use reverse pipetting techniques to insure bubbles are not created when dispensing the Read Buffer.

VIII Analysis of Results

analysis of results

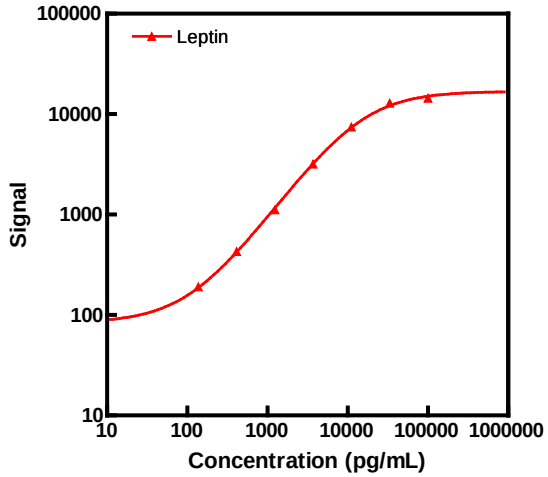
The Calibrators should be run in duplicate to generate a standard curve. The standard curve is modeled using least squares fitting algorithms so that signals from samples with known levels of the analyte of interest can be used to calculate the concentration of analyte in the sample. The assays have a wide dynamic range (3–4 logs) which allows accurate quantification in many samples without the need for dilution. The MSD DISCOVERY WORKBENCH® analysis software utilizes a 4-parameter logistic model (or sigmoidal dose-response) and includes a $1/Y^2$ weighting function. The weighting functionality is important because it provides a better fit of data over a wide dynamic range, particularly at the low end of the standard curve.

IX Typical Standard Curve

typical standard curve

The MSD Mouse Leptin Assay is designed for use with mouse serum and plasma samples.

The following standard curve is an example of the dynamic range of the assay. The actual signals may vary. A standard curve should be run for each set of samples and on each plate for the best quantification of unknown samples.



	Leptin	
Conc. (pg/mL)	**Average Signal**	**%CV**
0	83	12.1
137	191	3.5
412	429	3.9
1235	1119	9.0
3704	3185	2.5
11111	7454	3.5
33333	12900	4.0
100000	14418	5.6

X Sensitivity

sensitivity

The lower limit of detection (LLOD) is the calculated concentration of the signal that is 2.5 standard deviations over the zero Calibrator. The value below represents the average LLOD over multiple kit lots.

	Leptin
LLOD (pg/mL)	35

XI Spike Recovery

spike recovery

Serum, EDTA plasma, and heparin plasma were spiked with the Calibrators at multiple values throughout the range of the assay. The Calibrators were spiked into individual mouse samples and therefore spike recovery may depend on specific individual samples.
% Recovery = measured /expected x 100

Sample	Spike Level (pg/mL)	Conc. (pg/mL)	Conc. %CV	% Recovery
Spiked Serum	0	1721	4.8	
	20000	28636	9.1	107
	35000	48734	5.1	113
	70000	122817	2.0	153
Spiked EDTA Plasma	0	1592	2.1	
	20000	26918	3.9	101
	35000	42005	4.1	97
	70000	78602	2.1	98
Spiked Heparin Plasma	0	1473	0.6	
	20000	31699	2.9	120
	35000	50485	6.4	118
	70000	165466	0.9	206

XII Linearity

linearity

Linearity was measured by spiking Calibrator levels in pooled mouse samples followed by subsequent dilution.
Percent recovery is calculated as the measured concentration divided by the concentration of the previous dilution (expected).
% Recovery = measured x dilution factor / expected x 100

Sample	Fold Dilution	Conc. (pg/mL)	Conc. %CV	% Recovery
Serum	1	71548	3.1	
	2	64996	3.9	91
	4	61614	7.4	95
	8	60501	0.8	98
EDTA Plasma	1	83400	4.6	
	2	70095	2.4	84
	4	66197	3.8	97
	8	68477	3.0	103
Heparin Plasma	1	84224	1.4	
	2	68269	1.6	81
	4	66757	3.6	98
	8	69405	4.2	104

XIII Assay Components

assay components

Calibrator	
Analyte	Mouse leptin
Source	Purified, recombinant mouse leptin expressed in E. coli

Capture Antibody	
Analyte	Mouse leptin
Source	Rabbit polyclonal
Isoforms Recognized	Reacts with recombinant and natural mouse leptin
Species cross-reactivity	Mouse, rat

Detection Antibody	
Analyte	Mouse leptin
Source	Goat polyclonal
Isoforms Recognized	n/a
Species cross-reactivity	Mouse

XIV References

references

1. Matares G, Moschos S, Mantzoros CS. Leptin in Immunology. The Journal of Immunology, 2005 173: 3137–3142
2. Coll AP, Farooqi SI, O'Rahilly S. The Hormonal Control of Food Intake. 2007 Cell 129(2):l 252-262, 2007
3. Ahren B, Mansson S, Ginderich RL, Havel P. Regulation of plasma leptin in mice: influence of age, high-fat diet, and fasting. American Journal of Physiology. 1997 273(42): R113-R120

Summary Protocol

MSD 96-well MULTI-ARRAY Mouse Leptin Kit

MSD provides this summary protocol for your convenience.
Please read the entire detailed protocol prior to performing
the Mouse Leptin Assay.

Step 1 : Sample and Reagent Preparation

Bring all reagents to room temperature and thaw the Calibrator stock on ice.

Prepare Blocker A Solution.

Prepare serum or plasma samples.

Prepare an 8-point standard curve using supplied Calibrator:

- The Calibrator should be diluted in Diluent 100.
- Dilute the stock Calibrator 1:100 as indicated in Reagent Preparation section, then perform a series of 3-fold dilution steps and a no Calibrator blank.
- Diluted Calibrators should be kept on ice until use.

Note: *The standard curve can be modified as necessary to meet specific assay requirements.*

Prepare Detection Antibody Solution by diluting the 100X Anti-mLeptin Antibody to 1X in 5.0 mL of Diluent 17 per plate.

Prepare 20 mL of 1X Read Buffer T by diluting 4X Read Buffer T with deionized water.

Step 2 : Add Blocker A Solution

Dispense 150 µL/well Blocker A Solution.

Incubate at room temperature with vigorous shaking (300–1000 rpm) for 1 hour.

Step 3 : Wash and Add Sample or Calibrator

Wash plate 3 times with PBS-T.

Dispense 40 µL/well of 1X Detection Antibody Solution.

Immediately, dispense 10 µL/well Calibrator or Sample.

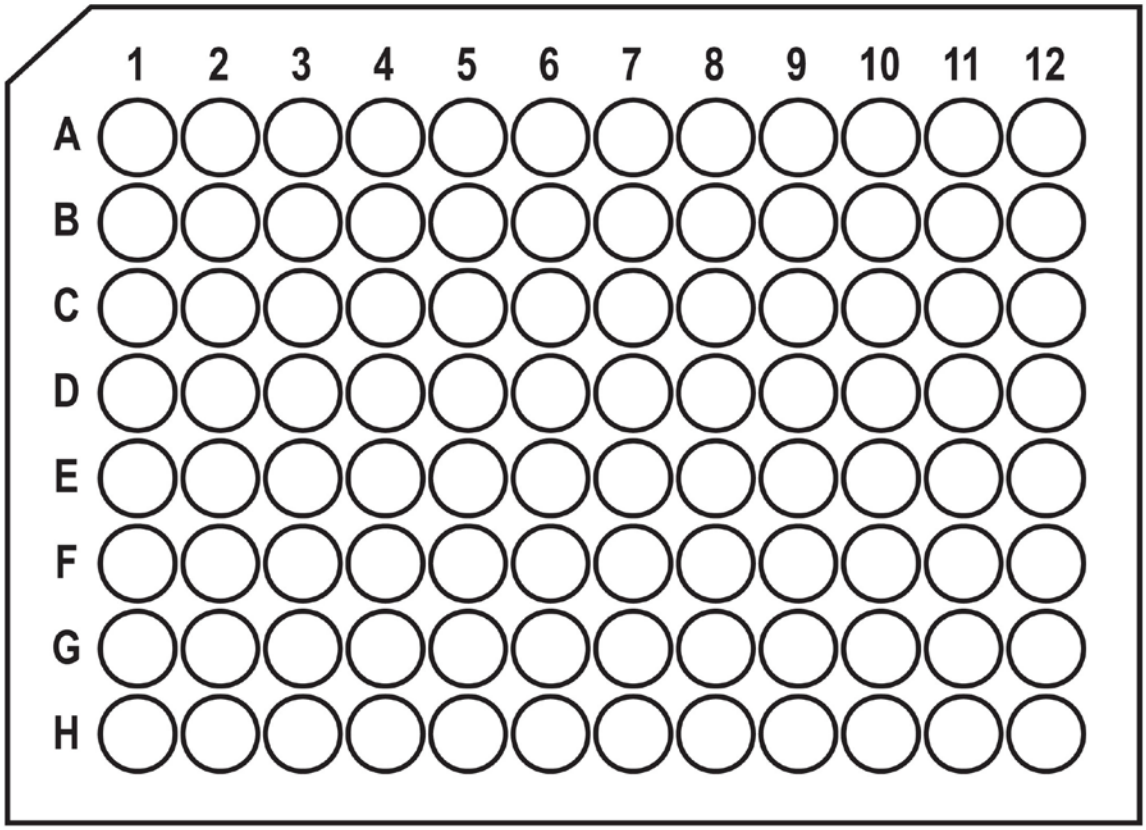
Incubate at room temperature with vigorous shaking (300–1000 rpm) for 2 hours.

Step 4 : Wash and Read Plate

Wash plate 3 times with PBS-T.

Dispense 150 µL/well 1X Read Buffer T.

Analyze plate on SECTOR instrument.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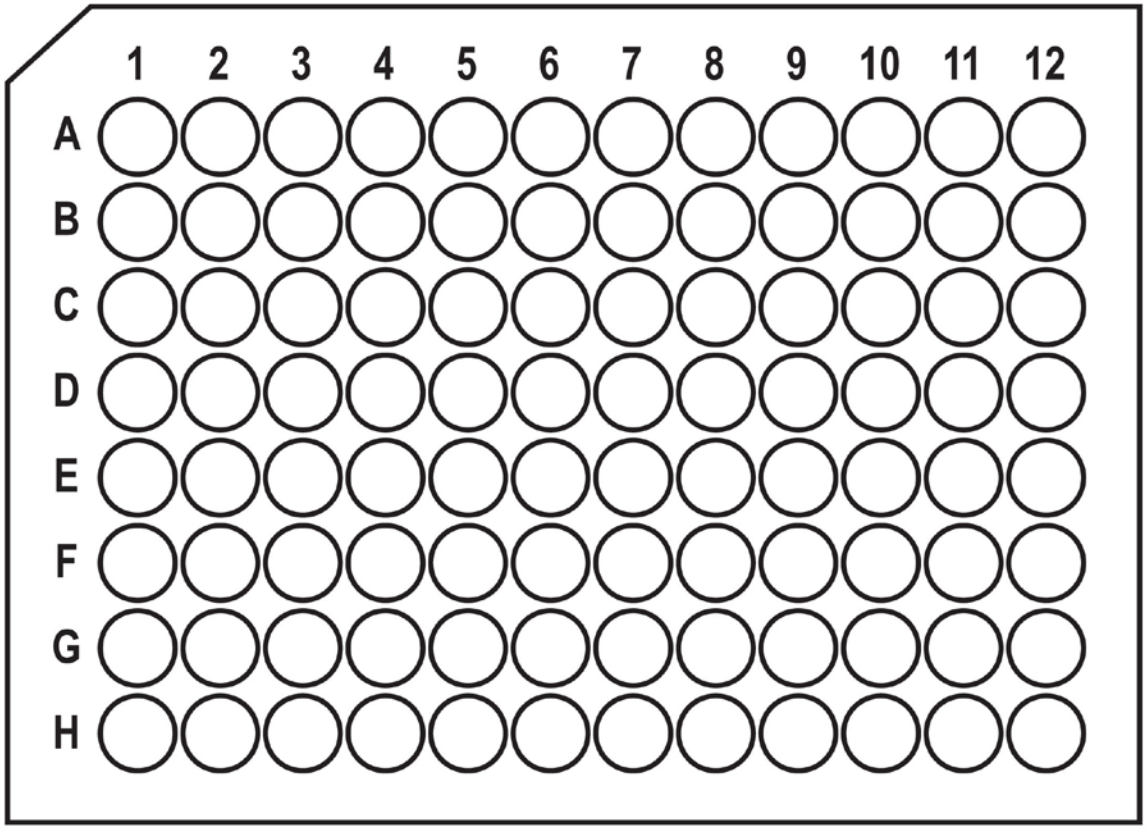
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