

# An ECL Multiplexed Assay for GPCRs

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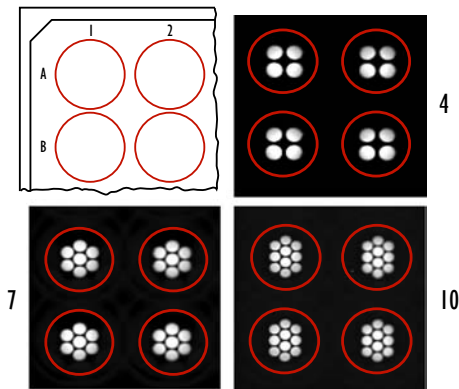


# An ECL Multiplexed Assay for GPCRs

## 1 Abstract

G Protein-Coupled Receptors (GPCRs) are integral membrane proteins that bind specific ligands and transmit changes in the external cellular environment to the interior of the cell. GPCRs constitute one of the largest families of proteins with the broadest span of therapeutic indications. They have proven to be highly tractable and selective drug targets with ~24% of the drug sales targeting GPCR-mediated mechanisms. Traditional screening of GPCRs has utilized the binding of a radioligand to a membrane receptor preparation, followed by separation of the unbound radioligand from the ligand-receptor complex. This gold standard method produces accurate and clean data, but is time-consuming and produces significant radioactive waste. Here, we present work done in conjunction with Meso Scale Discovery using their plate-based electrochemiluminescent technology to develop a 96-well multi-array, non-radioactive membrane receptor-ligand binding assay that simultaneously examines 2 related GPCRs within the same well (i.e. multiplexing).

## 2 Meso Scale Discovery Multiplexing



96 wells: 4, 7, and 10 spots/well

Multi-Spot™ plates enable multiplexing.

Microfluidics allows “addressing” to immobilize distinct species on each electrode.

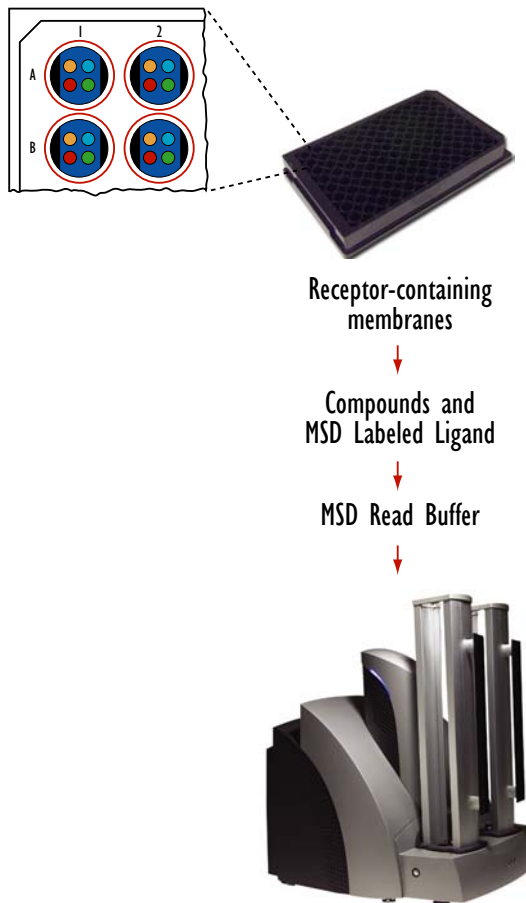
SECTOR™ Imager 6000 reader results  $\leq 0.05\%$  optical cross-talk.



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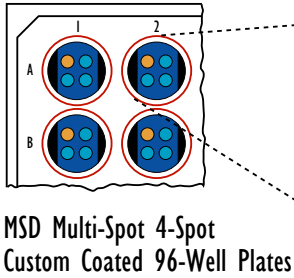
## 3 Assay Protocol for Multiplex GPCRs

1. Cellular membranes are immobilized onto 4 spatially distinct electrodes within the same well in MSD Multi-Spot 4-Spot Custom Coated 96-well plates. Dispense 0.2  $\mu\text{L}$  (0.5  $\mu\text{g}$  per spot) of membranes in Binding Buffer (25mM HEPES-KOH, pH 7.3, 1mM  $\text{CaCl}_2$ , 5mM  $\text{MgCl}_2$ , and protease inhibitors (Roche, 1836170)) and incubate for 1h at room temperature.
2. Wash plates with Binding Buffer.
3. Dispense 3  $\mu\text{L}$  of unlabeled compound, followed by 42  $\mu\text{L}$  MSD TAG labeled ligand in Binding Buffer and MSD Blocker A (0.5% final) and incubate for 1 hour at room temperature.
4. Wash plates with Binding Buffer.
5. Dispense 150  $\mu\text{L}$  of MSD Read Buffer T (2X, surfactant free) and image plate on an MSD SECTOR Imager 6000 reader. Plate is read in  $\sim 70$  seconds, regardless of plate type. Total time for the assay is 2.5 hours, including liquid handling time.

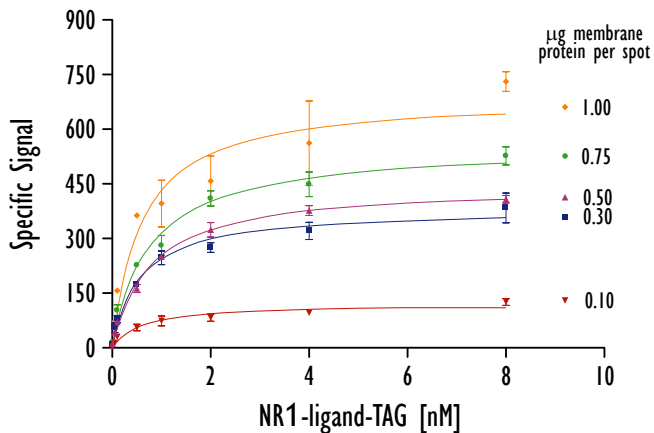
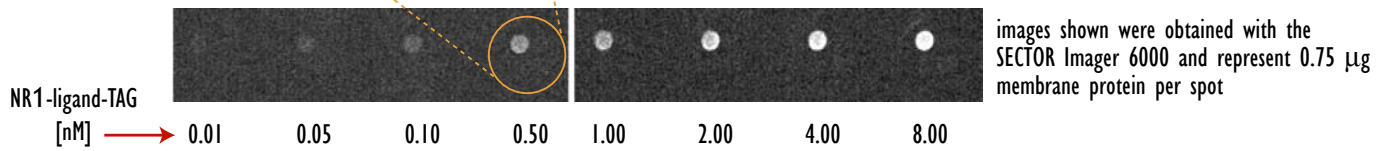


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## 4 Neuropeptide Receptor 1: Ligand & Membrane Titration



Neuropeptide Receptor 1 (NR1)-containing membranes were immobilized on one of the four spatially distinct electrodes, while CHO-K1 membranes prepared from the parent cell line (background) were immobilized on the remaining three electrodes within the same well. Titrations of membrane were assayed in ligand-binding isotherms.  $K_d$  values remain constant for each membrane concentration and agree with published data. Use of as little as 0.1  $\mu\text{g}$  of membrane protein per spot demonstrates the sensitivity of MSD Multi-Spot technology.

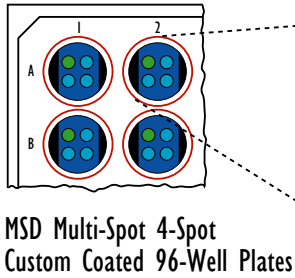


| $\mu\text{g}$ Membrane protein per spot | Signal/Background | $K_d$ , nM(95% C.I.) |
|---|-------------------|----------------------|
| 0.10                                    | 7.6               | 0.59 (0.14 - 1.00)   |
| 0.30                                    | 27.8              | 0.56 (0.28 - 0.85)   |
| 0.50                                    | 23.9              | 0.77 (0.58 - 0.97)   |
| .075                                    | 34.4              | 0.76 (0.41 - 1.11)   |
| 1.00                                    | 42.1              | 0.60 (0.10 - 1.09)   |

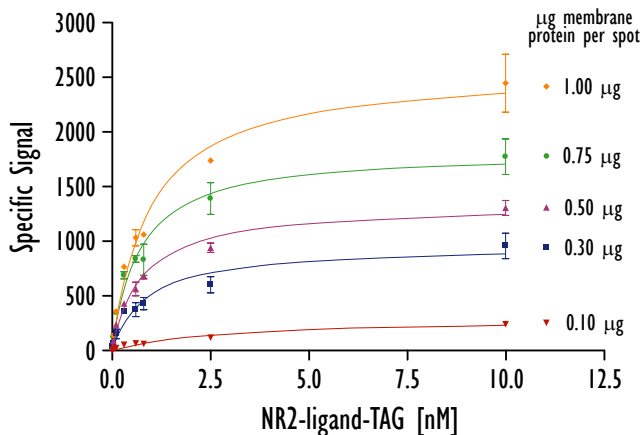
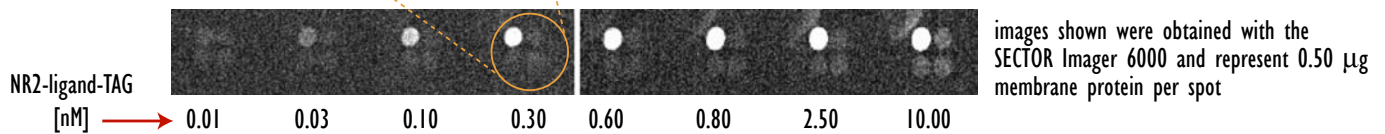
Background signal was obtained with membranes prepared from the parent cell line, CHO-K1.

# An ECL Multiplexed Assay for GPCRs

## 5 Neuropeptide Receptor 2: Ligand & Membrane Titration



Neuropeptide Receptor 2 (NR2)-containing membranes were immobilized on one of the four spatially distinct electrodes, while CHO-K1 membranes prepared from the parent cell line (background) were immobilized on the remaining three electrodes within the same well. The images show results obtained from a titration of the NR2 ligand labeled with MSD TAG, and agree with published data.

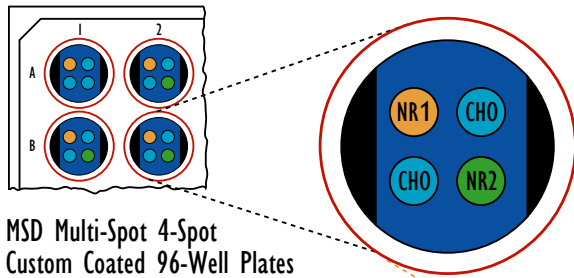


| µg Membrane protein per spot | Signal/Background | $K_d$ , nM(95% C.I.) |
|------------------------------|-------------------|----------------------|
| 0.10                         | 2.9               | 2.89 (0.87 - 4.91)   |
| 0.30                         | 11.6              | 0.87 (0.28 - 1.47)   |
| 0.50                         | 14.9              | 0.75 (0.46 - 1.04)   |
| .075                         | 21.3              | 0.69 (0.37 - 1.01)   |
| 1.00                         | 29.1              | 0.95 (0.58 - 1.32)   |

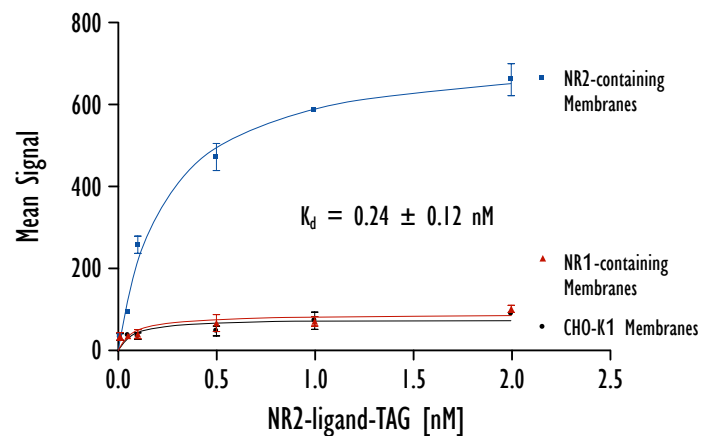
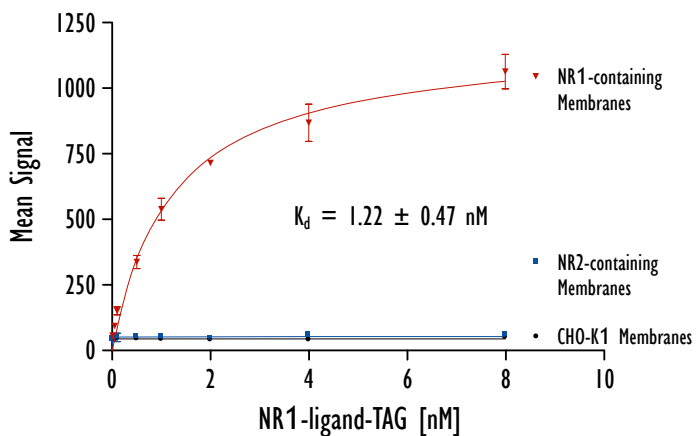
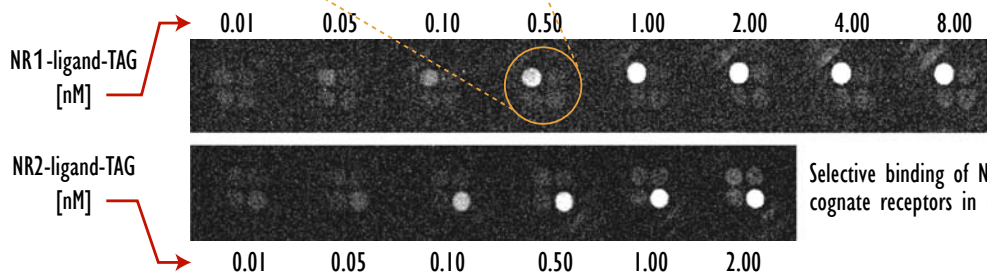
Background signal was obtained with membranes prepared from the parent cell line, CHO-K1.

# An ECL Multiplexed Assay for GPCRs

## 6 MSD Multiplex Assay with Neuropeptide Receptors 1 and 2

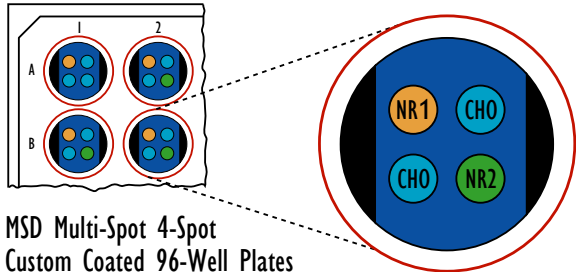


Neuropeptide Receptors 1 (NR1) and 2 (NR2) were immobilized on spatially distinct electrodes in the same well, 0.5  $\mu\text{g}$  per spot, in MSD Multi-Spot 4-Spot 96-well plates. Membranes isolated from the parent cell line, CHO-K1, were immobilized on the remaining two electrodes in each well. The  $K_d$  values obtained from these ligand-binding isotherms demonstrate that the binding specificity of individual labeled ligands was conserved in this multiplex assay.



# An ECL Multiplexed Assay for GPCRs

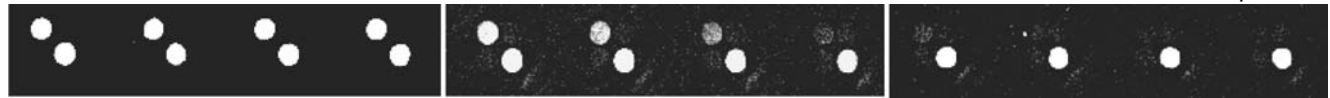
## 7 Multiplexed Neuropeptide Receptors: Selective Inhibition of NR1



0.5  $\mu$ g membrane per spot

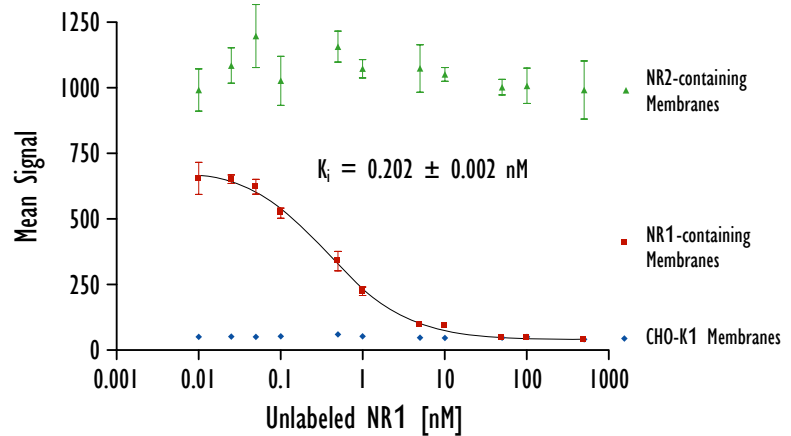
Neuropeptide Receptors 1 (NR1) and 2 (NR2), and CHO-K1 membranes prepared from the parent cell line, were immobilized on spatially distinct electrodes in the same well, 0.5  $\mu$ g per spot, in MSD Multi-Spot 4-Spot 96-well plates. Unlabeled NR1 ligand was titrated against 1 nM of MSD TAG-labeled NR1 ligand and 1 nM MSD TAG-labeled NR2 ligand. These results show that selective inhibition of neuropeptide receptor 1 can be achieved the the multiplex format.

NR1 is selectively inhibited



Increasing concentration of Unlabeled NR1 ligand

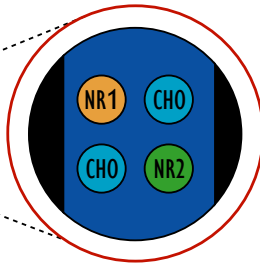
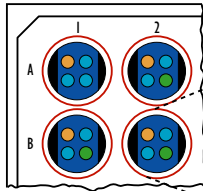
1 nM Labeled NR1 Ligand and 1 nM Labeled NR2 Ligand  
Challenged with Unlabeled NR1 Ligand



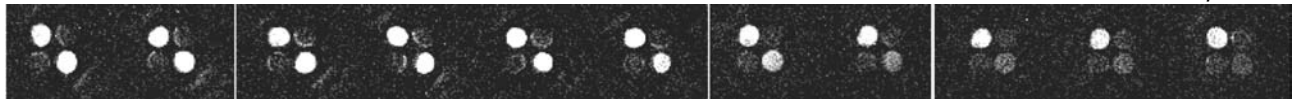
# An ECL Multiplexed Assay for GPCRs

## 8 Multiplexed Neuropeptide Receptors: Selective Inhibition of NR2

MSD Multi-Spot  
4-Spot Custom Coated  
96-Well Plates



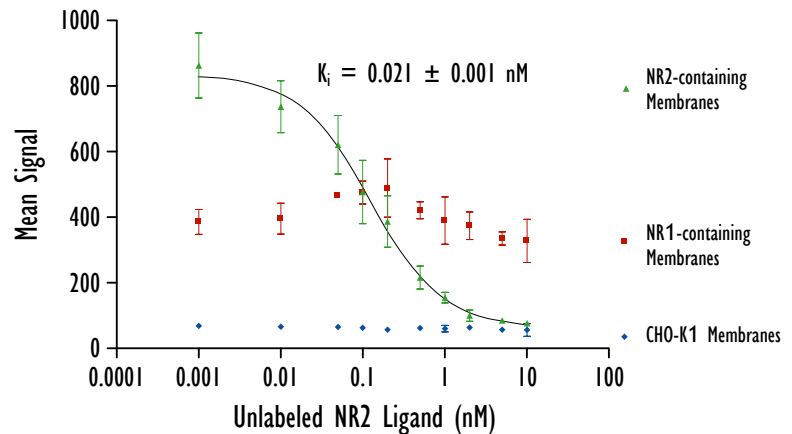
0.5  $\mu$ g membrane per spot



NR2 is selectively inhibited

Neuropeptide Receptors 1 (NR1) and 2 (NR2), and CHO-K1 membranes prepared from the parent cell line, were immobilized on spatially distinct electrodes in the same well, 0.5  $\mu$ g per spot, in MSD Multi-Spot 4-Spot 96-well plates. Unlabeled NR2 ligand was titrated against 1 nM of MSD TAG labeled NR1 ligand and 1 nM MSD TAG labeled NR2 ligand. These results show that neuropeptide receptor 2 can be achieved the the multiplex format.

1 nM Labeled NR1 Ligand and 1 nM Labeled NR2 Ligand  
Challenged with Unlabeled NR2 Ligand





# An ECL Multiplexed Assay for GPCRs

## 9 Validation of Pharmacology: Comparison of MSD with Filter Binding Assays

Neuropeptide 1 (NR1)-containing membranes, 1.0  $\mu\text{g}$  per well, were passively immobilized onto MSD Multi-Array Custom Coated 384-well plates. Competition studies were conducted in which 6 ligands or compounds were titrated against 1 nM MSD TAG-labeled NR1 ligand. Compounds were ranked based on potency and compared to analogous studies using Filter Binding, SPA, and LEADseeker™ technologies. Comparison of the results obtained with the MSD TAG-labeled ligand reveal equivalent performance to filter binding studies using radiolabeled ligand.

|                    | MSD<br>384-well       |            | Filter Binding<br>96-well |            | SPA<br>384-well       |            | LEADseeker<br>384-well** |            |
|--------------------|-----------------------|------------|---------------------------|------------|-----------------------|------------|--------------------------|------------|
|                    | IC <sub>50</sub> (nM) | Rank Order | IC <sub>50</sub> (nM)     | Rank Order | IC <sub>50</sub> (nM) | Rank Order | IC <sub>50</sub> (nM)    | Rank Order |
| NR1 ligand         | 0.7                   | 1          | 0.5                       | 1          | 0.3                   | 1          | 0.6                      | 1          |
| Synthetic Ligand 1 | 5                     | 2          | Not determined            |            | Not determined        |            | 37                       | 3          |
| NR1 ligand-YG5*    | 3                     | 2          | 2                         | 2          | 2                     | 2          | 4                        | 2          |
| Compound AMG 1     | 6                     | 2          | 23                        | 3          | 4                     | 2          | Not determined           |            |
| Synthetic Ligand 2 | 943                   | 3          | 580                       | 4          | 400                   | 3          | Not determined           |            |
| NR2 Ligand         | 11x10 <sup>3</sup>    | 4          | 18x10 <sup>3</sup>        | 5          | 75x10 <sup>3</sup>    | 3          | 25x10 <sup>3</sup>       | 4          |

\*NR1 ligand with the addition of 5 glycines and one tyrosine

\*\* Maximum inhibition observed with LEADseeker was 80% vs. 100% for others

## 10 Conclusions

Multiplex GPCR assays were achieved with Meso Scale Discovery's Multi-Spot 4-Spot Custom Coated 96-well plates and MSD TAG-labeled ligands. MSD TAG-labeled neuropeptide ligands exhibited pharmacological properties indistinguishable from radiolabeled ligands

Neuropeptide Receptors 1 and 2 immobilized in the same well (0.5  $\mu\text{g}$  per spot) exhibited conserved binding specificity with respect to MSD TAG-labeled ligands, and were selectively inhibited with the appropriate unlabeled ligands.

The assay is fast (complete in 2.5 hours) and sensitive, using as little as 0.1  $\mu\text{g}$  membrane per spot.

This assay is completely automated and affords a multiplex high-throughput workflow with the ability to screen 15,000 compounds per shift in workstation mode.